

ENHANCEMENT OF GROWTH IN PLANTS

TAB A 1 This application¹ claims the benefit of U.S. Provisional Patent Application Serial No. 60/036,048, filed January 27, 1997.

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FIELD OF THE INVENTION

10 The present invention relates to the
enhancement of growth in plants.

BACKGROUND OF THE INVENTION

15 The improvement of plant growth by the
application of organic fertilizers has been known and
carried out for centuries (H. Marschner, "Mineral
Nutrition of Higher Plants," Academic Press: New York
pg. 674 (1986). Modern man has developed a complex
20 inorganic fertilizer production system to produce an easy
product that growers and farmers can apply to soils or
growing crops to improve performance by way of growth
enhancement. Plant size, coloration, maturation, and
yield may all be improved by the application of
25 fertilizer products. Inorganic fertilizers include such
commonly applied chemicals as ammonium nitrate. Organic
fertilizers may include animal manures and composted lawn
debris, among many other sources.

In most recent years, researchers have sought
30 to improve plant growth through the use of biological
products. Insect and disease control agents such as
Beauveria bassiana and *Trichoderma harizamum* have been
registered for the control of insect and disease problems
and thereby indirectly improve plant growth and
35 performance (Fravel et al., "Formulation of

Microorganisms to Control Plant Diseases," Formulation of Microbial Biopesticides, Beneficial Microorganisms, and Nematodes, H.D. Burges, ed. Chapman and Hall: London (1996).

- 5 There is some indication of direct plant growth enhancement by way of microbial application or microbial by-products. Nodulating bacteria have been added to seeds of leguminous crops when introduced to a new site (Weaver et al., "Rhizobium," Methods of Soil Analysis, Part 2, Chemical and Microbiological Properties, 2nd ed., American Society of Agronomy: Madison (1982)). These bacteria may improve the nodulation efficiency of the plant and thereby improve the plant's ability to convert free nitrogen into a usable form, a process called
- 10 nitrogen fixation. Non-leguminous crops do not, as a rule, benefit from such treatment. Added bacteria such as *Rhizobium* directly parasitize the root hairs, then begin a mutualistic relationship by providing benefit to the plant while receiving protection and sustenance.
- 15 Mycorrhizal fungi have also been recognized as necessary microorganisms for optional growth of many crops, especially conifers in nutrient-depleted soils. Mechanisms including biosynthesis of plant hormones (Frankenberger et al., "Biosynthesis of Indole-3-Acetic
- 20 Acid by the Pine Ectomycorrhizal Fungus *Pisolithus tinctorius*," Appl. Environ. Microbiol. 53:2908-13 (1987)), increased uptake of minerals (Harley et al., "The Uptake of Phosphate by Excised Mycorrhizal Roots of Beech," New Phytologist 49:388-97 (1950) and Harley et
- 25 al., "The Uptake of Phosphate by Excised Mycorrhizal Roots of Beech. IV. The Effect of Oxygen Concentration Upon Host and Fungus," New Phytologist 52:124-32 (1953)), and water (A.B. Hatch, "The Physical Basis of Mycotrophy in *Pinus*," Black Rock Forest Bull. No. 6, 168 pp. (1937))
- 30 have been postulated. Mycorrhizal fungi have not

achieved the common frequency of use that nodulating bacteria have due to variable and inconsistent results with any given mycorrhizal strain and the difficulty of study of the organisms.

- 5 Plant growth-promoting rhizobacteria ("PGPR") have been recognized in recent years for improving plant growth and development. Hypothetical mechanisms range from direct influences (e.g., increased nutrient uptake) to indirect mechanisms (e.g., pathogen displacement).
- 10 Growth enhancement by application of a PGPR generally refers to inoculation with a live bacterium to the root system and achieving improved growth through bacterium-produced hormonal effects, siderophores, or by prevention of disease through antibiotic production, or competition.
- 15 In all of the above cases, the result is effected through root colonization, sometimes through the application of seed coatings. There is limited information to suggest that some PGPR strains may be direct growth promoters that enhance root elongation under gnotobiotic conditions
- 20 (Anderson et al., "Responses of Bean to Root Colonization With *Pseudomonas putida* in a Hydroponic System," Phytopathology 75:992-95 (1985), Lifshitz et al., "Growth Promotion of Canola (rapeseed) Seedlings by a Strain of *Pseudomonas putida* Under Gnotobiotic Conditions," Can. J. Microbiol. 33:390-95 (1987), Young et al., "PGPR: Is
- 25 There Relationship Between Plant Growth Regulators and the Stimulation of Plant Growth or Biological Activity?," Promoting Rhizobacteria: Progress and Prospects, Second International Workshop on Plant Growth-promoting
- 30 Rhizobacteria, pp. 182-86 (1991), Loper et al., "Influence of Bacterial Sources of Indole-3-Acetic Acid on Root Elongation of Sugar Beet," Phytopathology 76:386-89 (1986), and Müller et al., "Hormonal Interactions in the Rhizosphere of Maize (*Zea mays* L.) and Their Effect
- 35 on Plant Development," Z. Pflanzenernährung Bodenkunde

- 152:247-54 (1989); however, the production of plant growth regulators has been proposed as the mechanism mediating these effects. Many bacteria produce various plant growth regulators *in vitro* (Atzorn et al.,
- 5 "Production of Gibberellins and Indole-3-Acetic Acid by *Rhizobium phaseoli* in Relation to Nodulation of *Phaseolus vulgaris* Roots," Planta 175:532-38 (1988) and M. E. Brown, "Plant Growth Substances Produced by Micro-
- 10 35:443-51 (1972)) or antibiotics (Gardner et al., "Growth Promotion and Inhibition by Antibiotic-Producing Fluorescent *Pseudomonads* on Citrus Roots," Plant Soil 77:103-13 (1984)). Siderophore production is another mechanism proposed for some PGPR strains (Ahl et al.,
- 15 "Iron Bound-Siderophores, Cyanic Acid, and Antibiotics Involved in Suppression of *Thievaliopsis basicola* by a *Pseudomonas fluorescens* Strain," J. Phytopathol. 116:121-34 (1986), Kloepper et al., "Enhanced Plant Growth by Siderophores Produced by Plant Growth-Promoting
- 20 Rhizobacteria," Nature 286:885-86 (1980), and Kloepper et al., "*Pseudomonas siderophores*: A Mechanism Explaining Disease-Suppressive Soils," Curr. Microbiol. 4:317-20 (1980)). The colonization of root surfaces and thus the direct competition with pathogenic bacteria on the
- 25 surfaces is another mechanism of action (Kloepper et al., "Relationship of *in vitro* Antibiosis of Plant Growth-Promoting Rhizobacteria to Plant Growth and the Displacement of Root Microflora," Phytopathology 71:1020-24 (1981), Weller, et al., "Increased Growth of Wheat by
- 30 Seed Treatments With Fluorescent *Pseudomonads*, and Implications of *Pythium* Control," Can. J. Microbiol. 8:328-34 (1986), and Suslow et al., "Rhizobacteria of Sugar Beets: Effects of Seed Application and Root Colonization on Yield," Phytopathology 72:199-206
- 35 (1982)). Canola (rapeseed) studies have indicated PGPR

increased plant growth parameters including yields, seedling emergence and vigor, early-season plant growth (number of leaves and length of main runner), and leaf area (Kloepper et al., "Plant Growth-Promoting Rhizobacteria on Canola (rapeseed)," Plant Disease 72:42-46 (1988)). Studies with potato indicated greater yields when *Pseudomonas* strains were applied to seed potatoes (Burr et al., "Increased Potato Yields by Treatment of Seed Pieces With Specific Strains of *Pseudomonas* *Fluorescens* and *P. putida*," Phytopathology 68:1377-83 (1978), Kloepper et al., "Effect of Seed Piece Inoculation With Plant Growth-Promoting Rhizobacteria on Populations of *Erwinia carotovora* on Potato Roots and in Daughter Tubers," Phytopathology 73:217-19 (1983), Geels et al., "Reduction of Yield Depressions in High Frequency Potato Cropping Soil After Seed Tuber Treatments With Antagonistic Fluorescent *Pseudomonas* spp.," Phytopathol. Z. 108:207-38 (1983), Howie et al., "Rhizobacteria: Influence of Cultivar and Soil Type on Plant Growth and Yield of Potato," Soil Biol. Biochem. 15:127-32 (1983), and Vransy et al., "Growth and Yield of Potato Plants Inoculated With Rhizosphere Bacteria," Folia Microbiol. 29:248-53 (1984)). Yield increase was apparently due to the competitive effects of the PGPR to eliminate pathogenic bacteria on the seed tuber, possibly by antibiosis (Kloepper et al., "Effect of Seed Piece Inoculation With Plant Growth-Promoting Rhizobacteria on Populations of *Erwinia carotovora* on Potato Roots and in Daughter Tubers," Phytopathology 73:217-19 (1983), Kloepper et al., "Effects of Rhizosphere Colonization by Plant Growth-Promoting Rhizobacteria on Potato Plant Development and Yield," Phytopathology 70:1078-82 (1980), Kloepper et al., "Emergence-Promoting Rhizobacteria: Description and Implications for Agriculture," pp. 155-164, Iron, Siderophores, and Plant Disease, T.R.

- Swinburne, ed. Plenum, New York (1986), and Kloepper et al., "Relationship of *in vitro* Antibiosis of Plant Growth-Promoting Rhizobacteria to Plant Growth and the Displacement of Root Microflora," Phytopathology 71:1020-24 (1981)). In several studies, plant emergence was improved using PGPR (Tipping et al., "Development of Emergence-Promoting Rhizobacteria for Supersweet Corn," Phytopathology 76:938-41 (1990) (abstract) and Kloepper et al., "Emergence-Promoting Rhizobacteria: Description and Implications for Agriculture," pp. 155-164, Iron, Siderophores, and Plant Disease, T.R. Swinburne, ed. Plenum, New York (1986)). Numerous other studies indicated improved plant health upon treatment with rhizobacteria, due to biocontrol of plant pathogens (B. Schippers, "Biological Control of Pathogens With Rhizobacteria," Phil. Trans. R. Soc. Lond. B. 318:283-93 (1988), Schroth et al., "Disease-Suppressive Soil and Root-Colonizing Bacteria," Science 216:1376-81 (1982), Stutz et al., "Naturally Occurring Fluorescent Pseudomonads Involved in Suppression of Black Root Rot of Tobacco," Phytopathology 76:181-85 (1986), and D.M. Weller, "Biological Control of Soilborne Plant Pathogens in the Rhizosphere With Bacteria," Annu. Rev. Phytopathol. 26:379-407 (1988)).
- Pathogen-induced immunization of a plant has been found to promote growth. Injection of *Peronospora tabacina* externally to tobacco xylem not only alleviated stunting but also promoted growth and development. Immunized tobacco plants, in both greenhouse and field experiments, were approximately 40% taller, had a 40% increase in dry weight, a 30% increase in fresh weight, and 4-6 more leaves than control plants (Tuzun, S., et al., "The Effect of Stem Injection with *Peronospora tabacina* and Metalaxyl Treatment on Growth of Tobacco and Protection Against Blue Mould in the Field,"

Phytopathology, 74:804 (1984). These plants flowered approximately 2-3 weeks earlier than control plants (Tuzun, S., et al., "Movement of a Factor in Tobacco Infected with *Peronospora tabacina* Adam which Systemically Protects Against Blue Mould," Physiological Plant Pathology, 26:321-30 (1985)).

The present invention is directed to an improvement over prior plant growth enhancement procedures.

10

SUMMARY OF THE INVENTION

The present invention relates to a method of enhancing growth in plants. This method involves applying a hypersensitive response elicitor polypeptide or protein in a non-infectious form to plants or plant seeds under conditions to impart enhanced growth to the plants or to plants grown from the plant seeds.

As an alternative to applying a hypersensitive response elicitor polypeptide or protein to plants or plant seeds in order to impart enhanced growth to the plants or to plants grown from the seeds, transgenic plants or plant seeds can be utilized. When utilizing transgenic plants, this involves providing a transgenic plant transformed with a DNA molecule encoding a hypersensitive response elicitor polypeptide or protein and growing the plant under conditions effective to permit that DNA molecule to enhance growth. Alternatively, a transgenic plant seed transformed with a DNA molecule encoding a hypersensitive response elicitor polypeptide or protein can be provided and planted in soil. A plant is then propagated from the planted seed under conditions effective to permit that DNA molecule to enhance growth.

The present invention is directed to effecting any form of plant growth enhancement or promotion. This can occur as early as when plant growth begins from seeds or later in the life of a plant. For example, plant growth according to the present invention encompasses greater yield, increased quantity of seeds produced, increased percentage of seeds germinated, increased plant size, greater biomass, more and bigger fruit, earlier fruit coloration, and earlier fruit and plant maturation. As a result, the present invention provides significant economic benefit to growers. For example, early germination and early maturation permit crops to be grown in areas where short growing seasons would otherwise preclude their growth in that locale. Increased percentage of seed germination results in improved crop stands and more efficient seed use. Greater yield, increased size, and enhanced biomass production allow greater revenue generation from a given plot of land. It is thus apparent that the present invention constitutes a significant advance in agricultural efficiency.

BRIEF DESCRIPTION OF THE DRAWINGS

Figure 1 is a map of plasmid vector pCPP2139 which contains the *Erwinia amylovora* hypersensitive response elicitor gene.

Figure 2 is a map of plasmid vector pCPP50 which does not contain the *Erwinia amylovora* hypersensitive response elicitor gene but is otherwise the same as plasmid vector pCPP2139 shown in Figure 1. See Masui, et al., Bio/Technology 2:81-85 (1984), which is hereby incorporated by reference.

DETAILED DESCRIPTION OF THE INVENTION

The present invention relates to a method of enhancing growth in plants. This method involves
5 applying a hypersensitive response elicitor polypeptide or protein in a non-infectious form to all or part of a plant or a plant seed under conditions to impart enhanced growth to the plant or to a plant grown from the plant seed. Alternatively, plants can be treated in this
10 manner to produce seeds, which when planted, impart enhanced growth in progeny plants.

As an alternative to applying a hypersensitive response elicitor polypeptide or protein to plants or plant seeds in order to impart enhanced growth to the
15 plants or to plants grown from the seeds, transgenic plants or plant seeds can be utilized. When utilizing transgenic plants, this involves providing a transgenic plant transformed with a DNA molecule encoding a hypersensitive response elicitor polypeptide or protein
20 and growing the plant under conditions effective to permit that DNA molecule to enhance growth. Alternatively, a transgenic plant seed transformed with a DNA molecule encoding a hypersensitive response elicitor polypeptide or protein can be provided and planted in
25 soil. A plant is then propagated from the planted seed under conditions effective to permit that DNA molecule to enhance growth.

The hypersensitive response elicitor polypeptide or protein utilized in the present invention
30 can correspond to hypersensitive response elicitor polypeptides or proteins derived from a wide variety of fungal and bacterial pathogens. Such polypeptides or proteins are able to elicit local necrosis in plant tissue contacted by the elicitor.

Examples of suitable bacterial sources of polypeptide or protein elicitors include *Erwinia*, *Pseudomonas*, and *Xanthomonas* species (e.g., the following bacteria: *Erwinia amylovora*, *Erwinia chrysanthemi*,
5 *Erwinia stewartii*, *Erwinia carotovora*, *Pseudomonas syringae*, *Pseudomonas solanacearum*, *Xanthomonas campestris*, and mixtures thereof).

An example of a fungal source of a hypersensitive response elicitor protein or polypeptide
10 is *Phytophthora*. Suitable species of *Phytophthora* include *Phytophthora pythium*, *Phytophthora cryptogea*, *Phytophthora cinnamomi*, *Phytophthora capsici*, *Phytophthora megasperma*, and *Phytophthora citrophthora*.

The embodiment of the present invention where
15 the hypersensitive response elicitor polypeptide or protein is applied to the plant or plant seed can be carried out in a number of ways, including: 1) application of an isolated elicitor polypeptide or protein; 2) application of bacteria which do not cause
20 disease and are transformed with genes encoding a hypersensitive response elicitor polypeptide or protein; and 3) application of bacteria which cause disease in some plant species (but not in those to which they are applied) and naturally contain a gene encoding the
25 hypersensitive response elicitor polypeptide or protein. In addition, seeds in accordance with the present invention can be recovered from plants which have been treated with a hypersensitive response elicitor protein or polypeptide in accordance with the present invention.

30 In one embodiment of the present invention, the hypersensitive response elicitor polypeptides or proteins can be isolated from their corresponding organisms and applied to plants or plant seeds. Such isolation procedures are well known, as described in
35 Arlat, M., F. Van Gijsegem, J. C. Huet, J. C. Pemollet,

and C. A. Boucher, "PopA1, a Protein which Induces a Hypersensitive-like Response in Specific Petunia Genotypes is Secreted via the Hrp Pathway of *Pseudomonas solanacearum*," EMBO J. 13:543-553 (1994); He, S. Y., H. C. Huang, and A. Collmer, "*Pseudomonas syringae* pv. *syringae* Harpin_{PSS}: a Protein that is Secreted via the Hrp Pathway and Elicits the Hypersensitive Response in Plants," Cell 73:1255-1266 (1993); and Wei, Z.-M., R. J. Laby, C. H. Zumoff, D. W. Bauer, S.-Y. He, A. Collmer, and S. V. Beer, "Harpin Elicitor of the Hypersensitive Response Produced by the Plant Pathogen *Erwinia amylovora*," Science 257:85-88 (1992), which are hereby incorporated by reference. See also pending U.S. Patent Application Serial Nos. 08/200,024 and 08/062,024, which are hereby incorporated by reference. Preferably, however, the isolated hypersensitive response elicitor polypeptides or proteins of the present invention are produced recombinantly and purified as described below.

In other embodiments of the present invention, the hypersensitive response elicitor polypeptide or protein of the present invention can be applied to plants or plant seeds by applying bacteria containing genes encoding the hypersensitive response elicitor polypeptide or protein. Such bacteria must be capable of secreting or exporting the polypeptide or protein so that the elicitor can contact plant or plant seeds cells. In these embodiments, the hypersensitive response elicitor polypeptide or protein is produced by the bacteria in planta or on seeds or just prior to introduction of the bacteria to the plants or plant seeds.

In one embodiment of the bacterial application mode of the present invention, the bacteria do not cause the disease and have been transformed (e.g., recombinantly) with genes encoding a hypersensitive response elicitor polypeptide or protein. For example,

E. coli, which does not elicit a hypersensitive response in plants, can be transformed with genes encoding a hypersensitive response elicitor polypeptide or protein and then applied to plants. Bacterial species other than
5 *E. coli* can also be used in this embodiment of the present invention.

In another embodiment of the bacterial application mode of the present invention, the bacteria do cause disease and naturally contain a gene encoding a
10 hypersensitive response elicitor polypeptide or protein. Examples of such bacteria are noted above. However, in this embodiment, these bacteria are applied to plants or their seeds which are not susceptible to the disease carried by the bacteria. For example, *Erwinia amylovora*
15 causes disease in apple or pear but not in tomato. However, such bacteria will elicit a hypersensitive response in tomato. Accordingly, in accordance with this embodiment of the present invention, *Erwinia amylovora* can be applied to tomato plants or seeds to enhance
20 growth without causing disease in that species.

The hypersensitive response elicitor polypeptide or protein from *Erwinia chrysanthemi* has an amino acid sequence corresponding to SEQ. ID. No. 1 as follows:

25

Met Gln Ile Thr Ile Lys Ala His Ile Gly Gly Asp Leu Gly Val Ser
1 5 10 15

30

Gly Leu Gly Ala Gln Gly Leu Lys Gly Leu Asn Ser Ala Ala Ser Ser
20 25 30

Leu Gly Ser Ser Val Asp Lys Leu Ser Ser Thr Ile Asp Lys Leu Thr
35 40 45

35

Ser Ala Leu Thr Ser Met Met Phe Gly Gly Ala Leu Ala Gln Gly Leu
50 55 60

40

Gly Ala Ser Ser Lys Gly Leu Gly Met Ser Asn Gln Leu Gly Gln Ser
65 70 75 80

Phe Gly Asn Gly Ala Gln Gly Ala Ser Asn Leu Leu Ser Val Pro Lys
85 90 95

Ser Gly Gly Asp Ala Leu Ser Lys Met Phe Asp Lys Ala Leu Asp Asp
 100 105 110
 5 Leu Leu Gly His Asp Thr Val Thr Lys Leu Thr Asn Gln Ser Asn Gln
 115 120 125
 Leu Ala Asn Ser Met Leu Asn Ala Ser Gln Met Thr Gln Gly Asn Met
 130 135 140
 10 Asn Ala Phe Gly Ser Gly Val Asn Asn Ala Leu Ser Ser Ile Leu Gly
 145 150 155 160
 Asn Gly Leu Gly Gln Ser Met Ser Gly Phe Ser Gln Pro Ser Leu Gly
 165 170 175
 15 Ala Gly Gly Leu Gln Gly Leu Ser Gly Ala Gly Ala Phe Asn Gln Leu
 180 185 190
 Gly Asn Ala Ile Gly Met Gly Val Gly Gln Asn Ala Ala Leu Ser Ala
 195 200 205
 20 Leu Ser Asn Val Ser Thr His Val Asp Gly Asn Asn Arg His Phe Val
 210 215 220
 25 Asp Lys Glu Asp Arg Gly Met Ala Lys Glu Ile Gly Gln Phe Met Asp
 225 230 235 240
 Gln Tyr Pro Glu Ile Phe Gly Lys Pro Glu Tyr Gln Lys Asp Gly Trp
 245 250 255
 30 Ser Ser Pro Lys Thr Asp Asp Lys Ser Trp Ala Lys Ala Leu Ser Lys
 260 265 270
 Pro Asp Asp Asp Gly Met Thr Gly Ala Ser Met Asp Lys Phe Arg Gln
 275 280 285
 35 Ala Met Gly Met Ile Lys Ser Ala Val Ala Gly Asp Thr Gly Asn Thr
 290 295 300
 40 Asn Leu Asn Leu Arg Gly Ala Gly Gly Ala Ser Leu Gly Ile Asp Ala
 305 310 315 320
 Ala Val Val Gly Asp Lys Ile Ala Asn Met Ser Leu Gly Lys Leu Ala
 325 330 335
 45 Asn Ala

This hypersensitive response elicitor polypeptide or
 50 protein has a molecular weight of 34 kDa, is heat stable,
 has a glycine content of greater than 16%, and contains
 substantially no cysteine. The *Erwinia chrysanthemi*
 hypersensitive response elicitor polypeptide or protein
 is encoded by a DNA molecule having a nucleotide sequence
 55 corresponding to SEQ. ID. No. 2 as follows:

CGATTTTACC CGGGTGAACG TGCTATGACC GACAGCATCA CGGTATTCTGA CACCGTTACG 60

GCGTTTATGG CCGCGATGAA CCGGCATCAG GCGGCGCGCT GGTCGCCGCA ATCCGGCGTC 120

5 GATCTGGTAT TTCAGTTTGG GGACACCGGG CGTGAACCTA TGATGCAGAT TCAGCCGGGG 180

CAGCAATATC CCGGCATGTT GCGCACGCTG CTCGCTCGTC GTTATCAGCA GCGGCAGAG 240

TGCGATGGCT GCCATCTGTG CCTGAACGGC AGCGATGTAT TGATCCTCTG GTGGCCGCTG 300

10 CCGTCGGATC CCGGCAGTTA TCCGCAGGTG ATCGAACGTT TGTTTGAACCT GCGGGGAATG 360

ACGTTGCCGT CGCTATCCAT AGCACCGACG GCGCGTCCGC AGACAGGGAA CCGACGCGCC 420

15 CGATCATTA GATAAAGGCG GCTTTTTTTA TTGCAAAACG GTAACGGTGA GGAACCGTTT 480

CACCGTCGGC GTCACCTCAGT AACAGTATC CATCATGATG CCTACATCGG GATCGGCGTG 540

GGCATCCGTT GCAGATACTT TTGCGAACAC CTGACATGAA TGAGGAAACG AAATTATGCA 600

20 AATTACGATC AAAGCGCACA TCGGCGGTGA TTTGGGCGTC TCCGGTCTGG GGCTGGGTGC 660

TCAGGGACTG AAAGGACTGA ATTCCGCGGC TTCATCGCTG GGTTCAGCG TGATAAACT 720

25 GAGCAGCACC ATCGATAAGT TGACCTCCGC GCTGACTTCG ATGATGTTG GCGGCGCGCT 780

GGCGCAGGGG CTGGGCGCCA GCTCGAAGGG GCTGGGGATG AGCAATCAAC TGGGCCAGTC 840

TTTCGGCAAT GGCGCGCAGG GTGCGAGCAA CCTGCTATCC GTACCGAAAT CCGGCGGCGA 900

30 TGCGTTGTCA AAAATGTTTG ATAAAGCGCT GGACGATCTG CTGGGTCATG ACACCGTGAC 960

CAAGCTGACT AACCAGAGCA ACCAACTGGC TAATTCAATG CTGAACGCCA GCCAGATGAC 1020

35 CCAGGGTAAT ATGAATGCGT TCGGCAGCGG TGTGAACAAC GCACTGTCGT CCATTCTCGG 1080

CAACGGTCTC GGCCAGTCGA TGAGTGGCTT CTCTCAGCCT TCTCTGGGGG CAGGCGGCTT 1140

GCAGGGCCTG AGCGGCGCGG GTGCATTCAA CCAGTTGGGT AATGCCATCG GCATGGGCGT 1200

40 GGGGCAGAAT GCTGCGCTGA GTGCGTTGAG TAACGTCAGC ACCCAGTAG ACGSTAACAA 1260

CCGCCACTTT GTAGATAAAG AAGATCGCGG CATGGCGAAA GAGATCGGCC AGTTTATGGA 1320

45 TCAGTATCCG GAAATATTCT GTAAACCGGA ATACCAGAAA GATGGCTGGA GTTCGCCGAA 1380

GACGGACGAC AAATCCTGGG CTAAAGCGCT GAGTAAACCG GATGATGACG GTATGACCGG 1440

CGGCTGGATC CCGGCAGTTA TCCGCAGGTG ATCGAACGTT TGTTTGAACCT GCGGGGAATG

CGCCAGCATG GACAAATTCC GTCAGGCGAT GGGTATGATC AAAAGCGCGG TGGCGGGTGA 1500

TACCGGCAAT ACCAACCTGA ACCTGCGTGG CGCGGGCGGT GCATCGCTGG GTATCGATGC 1560

5 GGCTGTCGTC GGCGATAAAA TAGCCAACAT GTCGCTGGGT AAGCTGGCCA ACGCCTGATA 1620

ATCTGTGCTG GCCTGATAAA GCGGAAACGA AAAAAGAGAC GGGGAAGCCT GTCTCTTTTC 1680

TTATTATGCG GTTTATGCGG TTACCTGGAC CGGTTAATCA TCGTCATCGA TCTGGTACAA 1740

10 ACGCACATTT TCCCGTTCAT TCGCGTCGTT ACGCGCCACA ATCGCGATGG CATCTTCCTC 1800

GTCGCTCAGA TTGCGCGGCT GATGGGGAAC GCCGGGTGGA ATATAGAGAA ACTCGCCGGC 1860

15 CAGATGGAGA CACGTCTGCG ATAAATCTGT GCCGTAACGT GTTCTATCC GCCCCTTTAG 1920

CAGATAGATT GCGGTTTCGT AATCAACATG GTAATGCGGT TCCGCCTGTG CGCCGGCCGG 1980

GATCACCACA ATATTCATAG AAAGCTGTCT TGCACCTACC GTATCGCGGG AGATACCGAC 2040

20 AAAATAGGGC AGTTTTTGCG TGGTATCCGT GGGGTGTTC GGCCTGACAA TCTTGAGTTG 2100

GTCGTCATC ATCTTTCTCC ATCTGGGCGA CCTGATCGGT T 2141

25 The hypersensitive response elicitor polypeptide or protein derived from *Erwinia amylovora* has an amino acid sequence corresponding to SEQ. ID. No. 3 as follows:

30 Met Ser Leu Asn Thr Ser Gly Leu Gly Ala Ser Thr Met Gln Ile Ser
1 5 10 15

Ile Gly Gly Ala Gly Gly Asn Asn Gly Leu Leu Gly Thr Ser Arg Gln
20 25 30

35 Asn Ala Gly Leu Gly Gly Asn Ser Ala Leu Gly Leu Gly Gly Gly Asn
35 40 45

40 Gln Asn Asp Thr Val Asn Gln Leu Ala Gly Leu Leu Thr Gly Met Met
50 55 60

Met Met Met Ser Met Met Gly Gly Gly Gly Leu Met Gly Gly Gly Leu
65 70 75 80

45 Gly Gly Gly Leu Gly Asn Gly Leu Gly Gly Ser Gly Gly Leu Gly Glu
85 90 95

Gly Leu Ser Asn Ala Leu Asn Asp Met Leu Gly Gly Ser Leu Asn Thr
100 105 110

50

	Leu Gly Ser Lys Gly Gly Asn Asn Thr Thr Ser Thr Thr Asn Ser Pro	115	120	125
5	Leu Asp Gln Ala Leu Gly Ile Asn Ser Thr Ser Gln Asn Asp Asp Ser	130	135	140
	Thr Ser Gly Thr Asp Ser Thr Ser Asp Ser Ser Asp Pro Met Gln Gln	145	150	155
10	Leu Leu Lys Met Phe Ser Glu Ile Met Gln Ser Leu Phe Gly Asp Gly	165	170	175
	Gln Asp Gly Thr Gln Gly Ser Ser Ser Gly Gly Lys Gln Pro Thr Glu	180	185	190
15	Gly Glu Gln Asn Ala Tyr Lys Lys Gly Val Thr Asp Ala Leu Ser Gly	195	200	205
20	Leu Met Gly Asn Gly Leu Ser Gln Leu Leu Gly Asn Gly Gly Leu Gly	210	215	220
	Gly Gly Gln Gly Gly Asn Ala Gly Thr Gly Leu Asp Gly Ser Ser Leu	225	230	235
25	Gly Gly Lys Gly Leu Gln Asn Leu Ser Gly Pro Val Asp Tyr Gln Gln	245	250	255
	Leu Gly Asn Ala Val Gly Thr Gly Ile Gly Met Lys Ala Gly Ile Gln	260	265	270
30	Ala Leu Asn Asp Ile Gly Thr His Arg His Ser Ser Thr Arg Ser Phe	275	280	285
35	Val Asn Lys Gly Asp Arg Ala Met Ala Lys Glu Ile Gly Gln Phe Met	290	295	300
	Asp Gln Tyr Pro Glu Val Phe Gly Lys Pro Gln Tyr Gln Lys Gly Pro	305	310	315
40	Gly Gln Glu Val Lys Thr Asp Asp Lys Ser Trp Ala Lys Ala Leu Ser	325	330	335
	Lys Pro Asp Asp Asp Gly Met Thr Pro Ala Ser Met Glu Gln Phe Asn	340	345	350
45	Lys Ala Lys Gly Met Ile Lys Arg Pro Met Ala Gly Asp Thr Gly Asn	355	360	365
50	Gly Asn Leu Gln Ala Arg Gly Ala Gly Gly Ser Ser Leu Gly Ile Asp	370	375	380
	Ala Met Met Ala Gly Asp Ala Ile Asn Asn Met Ala Leu Gly Lys Leu	385	390	395
55	Gly Ala Ala			400

This hypersensitive response elicitor polypeptide or
 60 protein has a molecular weight of about 39 kDa, has a pI
 of approximately 4.3, and is heat stable at 100°C for at

least 10 minutes. This hypersensitive response elicitor polypeptide or protein has substantially no cysteine. The hypersensitive response elicitor polypeptide or protein derived from *Erwinia amylovora* is more fully described in Wei, Z.-M., R. J. Laby, C. H. Zumoff, D. W. Bauer, S.-Y. He, A. Collmer, and S. V. Beer, "Harpin, Elicitor of the Hypersensitive Response Produced by the Plant Pathogen *Erwinia amylovora*," Science 257:85-88 (1992), which is hereby incorporated by reference. The DNA molecule encoding this polypeptide or protein has a nucleotide sequence corresponding to SEQ. ID. No. 4 as follows:

15 AAGCTTCGGC ATGGCACGTT TGACCGTTGG GTCGGCAGGG TACGTTTGAA TTATTCATAA 60
GAGGAATACG TTATGAGTCT GAATACAAGT GGGCTGGGAG CGTCAACGAT GCAAATTTCT 120
ATCGGCGGTG CGGGCGGAAA TAACGGGTG CTGGGTACCA GTCGCCAGAA TGCTGGGTTG 180
20 GGTGGCAATT CTGCACTGGG GCTGGGCGGC GGTAATCAAA ATGATACCGT CAATCAGCTG 240
GCTGGCTTAC TCACCGGCAT GATGATGATG ATGAGCATGA TGGGCGGTGG TGGGCTGATG 300
25 GCGGGTGGCT TAGGCGGTGG CTTAGGTAAT GGCTTGGGTG GCTCAGGTGG CCTGGGCGAA 360
GGACTGTCTGA ACGCGCTGAA CGATATGTTA GCGGGTTCGC TGAACACGCT GGGCTCGAAA 420
GGCGGCAACA ATACCACTTC AACACAAAT TCCCCGCTGG ACCAGGCGCT GGGTATTAAC 480
30 TCAACGTCCC AAAACGACGA TTCCACCTCC GGCACAGATT CCACCTCAGA CTCCAGCGAC 540
CCGATGCAGC AGCTGCTGAA GATGTTTCAGC GAGATAATGC AAAGCCTGTT TGGTGATGGG 600
35 CAAGATGGCA CCCAGGGCAG TTCCTCTGGG GGCAAGCAGC CGACCGAAGG CGAGCAGAAC 660
GCCTATAAAA AAGGAGTCAC TGATGCGCTG TCGGGCCTGA TGGGTAATGG TCTGAGCCAG 720
CTCCTTGCCA ACGGGGGACT GGGAGGTGGT CAGGGCGGTA ATGCTGGCAC GGGTCTTGAC 780
40 GGTTCGTCGC TGGGCGGCAA AGGGCTGCAA AACCTGAGCG GGCCGGTGGA CTACCAGCAG 840
TTAGGTAAAC CCGTGGGTAC CGGTATCGGT ATGAAAGCGG GCATTCAGGC GCTGAATGAT 900
45 ATCGGTACGC ACAGGCACAG TTCAACCCGT TCTTTCGTCA ATAAAGGCGA TCGGGCGATG 960
GCGAAGGAAA TCGGTCAATT CATGGACCAG TATCCTGAGG TGTTTGGCAA GCCGCAGTAC 1020
CAGAAAGGCC CGGGTCAGGA GGTGAAAACC GATGACAAAT CATGGGCAAA AGCACTGAGC 1080
50 AAGCCAGATG ACGACGGAAT GACACCAGCC AGTATGGAGC AGTTCAACAA AGCCAAGGGC 1140
ATGATCAAAA GGCCCATGGC GGGTGATACC GGCAACGGCA ACCTGCAGGC ACGCGGTGCC 1200
55 GGTGGTTCTT CGCTGGGTAT TGATGCCATG ATGGCCGGTG ATGCCATTAA CAATATGGCA 1260
CTTGGAAGC TGGGCGCGGC TTAAGCTT 1288

The hypersensitive response elicitor polypeptide or protein derived from *Pseudomonas syringae* has an amino acid sequence corresponding to SEQ. ID. No. 5 as follows:

5 Met Gln Ser Leu Ser Leu Asn Ser Ser Ser Leu Gln Thr Pro Ala Met
1 5 10 15

10 Ala Leu Val Leu Val Arg Pro Glu Ala Glu Thr Thr Gly Ser Thr Ser
20 25 30

Ser Lys Ala Leu Gln Glu Val Val Val Lys Leu Ala Glu Glu Leu Met
35 40 45

15 Arg Asn Gly Gln Leu Asp Asp Ser Ser Pro Leu Gly Lys Leu Leu Ala
50 55 60

Lys Ser Met Ala Ala Asp Gly Lys Ala Gly Gly Gly Ile Glu Asp Val
65 70 75 80

20 Ile Ala Ala Leu Asp Lys Leu Ile His Glu Lys Leu Gly Asp Asn Phe
85 90 95

25 Gly Ala Ser Ala Asp Ser Ala Ser Gly Thr Gly Gln Gln Asp Leu Met
100 105 110

Thr Gln Val Leu Asn Gly Leu Ala Lys Ser Met Leu Asp Asp Leu Leu
115 120 125

30 Thr Lys Gln Asp Gly Gly Thr Ser Phe Ser Glu Asp Asp Met Pro Met
130 135 140

Leu Asn Lys Ile Ala Gln Phe Met Asp Asp Asn Pro Ala Gln Phe Pro
145 150 155 160

35 Lys Pro Asp Ser Gly Ser Trp Val Asn Glu Leu Lys Glu Asp Asn Phe
165 170 175

40 Leu Asp Gly Asp Glu Thr Ala Ala Phe Arg Ser Ala Leu Asp Ile Ile
180 185 190

Gly Gln Gln Leu Gly Asn Gln Gln Ser Asp Ala Gly Ser Leu Ala Gly
195 200 205

45 Thr Gly Gly Gly Leu Gly Thr Pro Ser Ser Phe Ser Asn Asn Ser Ser
210 215 220

Val Met Gly Asp Pro Leu Ile Asp Ala Asn Thr Gly Pro Gly Asp Ser
225 230 235 240

50 Gly Asn Thr Arg Gly Glu Ala Gly Gln Leu Ile Gly Glu Leu Ile Asp
245 250 255

Arg Gly Leu Gln Ser Val Leu Ala Gly Gly Gly Leu Gly Thr Pro Val
260 265 270

55 Asn Thr Pro Gln Thr Gly Thr Ser Ala Asn Gly Gly Gln Ser Ala Gln
275 280 285

Asp Leu Asp Gln Leu Leu Gly Gly Leu Leu Leu Lys Gly Leu Glu Ala
 290 295 300
 5 Thr Leu Lys Asp Ala Gly Gln Thr Gly Thr Asp Val Gln Ser Ser Ala
 305 310 315 320
 Ala Gln Ile Ala Thr Leu Leu Val Ser Thr Leu Leu Gln Gly Thr Arg
 325 330 335
 10 Asn Gln Ala Ala Ala
 340

This hypersensitive response elicitor polypeptide or
 15 protein has a molecular weight of 34-35 kDa. It is rich
 in glycine (about 13.5%) and lacks cysteine and tyrosine.
 Further information about the hypersensitive response
 elicitor derived from *Pseudomonas syringae* is found in
 He, S. Y., H. C. Huang, and A. Collmer, "*Pseudomonas*
 20 *syringae* pv. *syringae* Harpin_{pss}: a Protein that is
 Secreted via the Hrp Pathway and Elicits the
 Hypersensitive Response in Plants," *Cell* 73:1255-1266
 (1993), which is hereby incorporated by reference. The
 DNA molecule encoding the hypersensitive response
 25 elicitor from *Pseudomonas syringae* has a nucleotide
 sequence corresponding to SEQ. ID. No. 6 as follows:

ATGCAGAGTC TCAGTCTTAA CAGCAGCTCG CTGCAAACCC CGGCAATGGC CCTGTGCTCTG 60
 30 GTACGTCCTG AAGCCGAGAC GACTGGCAGT ACGTCGAGCA AGGCGCTTCA GGAAGTTGTC 120
 GTGAAGCTGG CCGAGGAAC TATGCGCAAT GGTCAACTCG ACGACAGCTC GCCATTGGGA 180
 35 AACTGTGTTG CCAAGTCGAT GGCCGAGAT GGCAAGGCGG GCGGCGGTAT TGAGGATGTC 240
 ATCGCTGCGC TGGACAAGCT GATCCATGAA AAGCTCGGTG ACAACTTCGG CGCGTCTGCG 300
 GACAGCGCCT CGGGTACCGG ACAGCAGGAC CTGATGACTC AGGTGCTCAA TGGCCTGGCC 360
 40 AAGTCGATGC TCGATGATCT TCTGACCAAG CAGGATGGCG GGACAAGCTT CTCCGAAGAC 420
 GATATGCCGA TGCTGAACAA GATCGCGCAG TTCATGGATG ACAATCCCGC ACAGTTTCCC 480
 AAGCCGGAAT CGGGCTCCTG GGTGAACGAA CTCAAGGAAG ACAACTTCCT TGATGGCGAC 540
 45 GAAACGGCTG CGTTCCGTTC GGCACGAC ATCATTGGCC AGCAACTGGG TAATCAGCAG 600
 AGTGACGCTG GCAGTCTGGC AGGGACGGGT GGAGGTCTGG GCACTCCGAG CAGTTTTC 660
 50 AACAACTCGT CCGTGATGGG TGATCCGCTG ATCGACGCCA ATACCGGTCC CGGTGACAGC 720
 GGCAATACCC GTGGTGAAGC GGGGCAACTG ATCGGCGAGC TTATCGACCG TGGCCTGCAA 780
 TCGGTATTGG CCGGTGGTGG ACTGGGCACA CCCGTAAACA CCGGCGAGAC CGGTACGTCG 840

GCGAATGGCG GACAGTCCGC TCAGGATCTT GATCAGTTGC TGGGCGGCTT GCTGCTCAAG 900
 GGCCTGGAGG CAACGCTCAA GGATGCCGGG CAAACAGGCA CCGACGTGCA GTCGAGCGCT 960
 5 GCGCAAATCG CCACCTTGCT GGTCACTACG CTGCTGCAAG GCACCCGCAA TCAGGCTGCA 1020
 GCCTGA 1026

10 The hypersensitive response elicitor
 polypeptide or protein derived from *Pseudomonas*
solanacearum has an amino acid sequence corresponding to
 SEQ. ID. No. 7 as follows:

15 Met Ser Val Gly Asn Ile Gln Ser Pro Ser Asn Leu Pro Gly Leu Gln
 1 5 10 15
 Asn Leu Asn Leu Asn Thr Asn Thr Asn Ser Gln Gln Ser Gly Gln Ser
 20 20 25 30
 Val Gln Asp Leu Ile Lys Gln Val Glu Lys Asp Ile Leu Asn Ile Ile
 35 40 45
 25 Ala Ala Leu Val Gln Lys Ala Ala Gln Ser Ala Gly Gly Asn Thr Gly
 50 55 60
 Asn Thr Gly Asn Ala Pro Ala Lys Asp Gly Asn Ala Asn Ala Gly Ala
 65 70 75 80
 30 Asn Asp Pro Ser Lys Asn Asp Pro Ser Lys Ser Gln Ala Pro Gln Ser
 85 90 95
 Ala Asn Lys Thr Gly Asn Val Asp Asp Ala Asn Asn Gln Asp Pro Met
 100 105 110
 35 Gln Ala Leu Met Gln Leu Leu Glu Asp Leu Val Lys Leu Leu Lys Ala
 115 120 125
 40 Ala Leu His Met Gln Gln Pro Gly Gly Asn Asp Lys Gly Asn Gly Val
 130 135 140
 Gly Gly Ala Asn Gly Ala Lys Gly Ala Gly Gly Gln Gly Gly Leu Ala
 145 150 155 160
 45 Glu Ala Leu Gln Glu Ile Glu Gln Ile Leu Ala Gln Leu Gly Gly Gly
 165 170 175
 Gly Ala Gly Ala Gly Gly Ala Gly Gly Gly Val Gly Gly Ala Gly Gly
 180 185 190
 50 Ala Asp Gly Gly Ser Gly Ala Gly Gly Ala Gly Gly Ala Asn Gly Ala
 195 200 205
 55 Asp Gly Gly Asn Gly Val Asn Gly Asn Gln Ala Asn Gly Pro Gln Asn
 210 215 220
 Ala Gly Asp Val Asn Gly Ala Asn Gly Ala Asp Asp Gly Ser Glu Asp
 225 230 235 240
 60

Gln Gly Gly Leu Thr Gly Val Leu Gln Lys Leu Met Lys Ile Leu Asn
245 250 255

5 Ala Leu Val Gln Met Met Gln Gln Gly Gly Leu Gly Gly Gly Asn Gln
260 265 270

Ala Gln Gly Gly Ser Lys Gly Ala Gly Asn Ala Ser Pro Ala Ser Gly
275 280 285

10 Ala Asn Pro Gly Ala Asn Gln Pro Gly Ser Ala Asp Asp Gln Ser Ser
290 295 300

Gly Gln Asn Asn Leu Gln Ser Gln Ile Met Asp Val Val Lys Glu Val
305 310 315 320

15 Val Gln Ile Leu Gln Gln Met Leu Ala Ala Gln Asn Gly Gly Ser Gln
325 330 335

Gln Ser Thr Ser Thr Gln Pro Met
340

20

It is encoded by a DNA molecule having a nucleotide
sequence corresponding SEQ. ID. No. 8 as follows:

25 ATGTCAGTCG GAAACATCCA GAGCCCGTCG AACCTCCCGG GTCTGCAGAA CCTGAACCTC 60

AACACCAACA CCAACAGCCA GCAATCGGGC CAGTCCGTGC AAGACCTGAT CAAGCAGGTC 120

30 GAGAAGGACA TCCTCAACAT CATCGCAGCC CTCGTGCAGA AGGCCGCACA GTCGGCGGGC 180

GGCAACACCG GTAACACCGG CAACGCGCCG GCGAAGGACG GCAATGCCAA CGCGGGCGCC 240

35 AACGACCCGA GCAAGAACGA CCCGAGCAAG AGCCAGGCTC CGCAGTCGGC CAACAAGACC 300

GGCAACGTCG ACGACGCCAA CAACCAGGAT CCGATGCAAG CGCTGATGCA GCTGCTGGAA 360

GACCTGGTGA AGCTGCTGAA GCGGCCCCTG CACATGCAGC AGCCCGGCGG CAATGACAAG 420

40 GGCAACGGCG TGGGCGGTGC CAACGGCGCC AAGGGTGCCG GCGGCCAGGG CGGCCTGGCC 480

GAAGCGCTGC AGGAGATCGA GCAGATCCTC GCCCAGCTCG GCGGCGGCGG TGCTGGCGCC 540

GGCGGCGCGG GTGGCGGTGT CGGCGGTGCT GGTGGCGCGG ATGGCGGCTC CGGTGCGGGT 600

45 GGCGCAGGCG GTGCGAACGG CGCCGACGGC GGCAATGGCG TGAACGGCAA CCAGGCGAAC 660

GGCCCGCAGA ACGCAGGCGA TGTCAACGGT GCCAACGGCG CGGATGACGG CAGCGAAGAC 720

50 CAGGGCGGCC TCACCGGCGT GCTGCAAAAG CTGATGAAGA TCCTGAACGC GCTGGTGCAG 780

ATGATGCAGC AAGGCGGCCT CGGCGGCGGC AACCAGGCGC AGGGCGGCTC GAAGGGTGCC 840

GGCAACGCCT CGCCGCTTC CGGCGCGAAC CCGGGCGCGA ACCAGCCCGG TTCGGCGGAT 900

55 GATCAATCGT CCGGCCAGAA CAATCTGCAA TCCCAGATCA TGGATGTGGT GAAGGAGGTC 960

GTCCAGATCC TGCAGCAGAT GCTGGCGGCG CAGAACGGCG GCAGCCAGCA GTCCACCTCG 1020

60 ACGCAGCCGA TGTA 1035

Further information regarding the hypersensitive response elicitor polypeptide or protein derived from *Pseudomonas solanacearum* is set forth in Arlat, M., F. Van Gijsegem, J. C. Huet, J. C. Pemollet, and C. A. Boucher, "PopA1, a
 5 Protein which Induces a Hypersensitive-like Response in Specific Petunia Genotypes, is Secreted via the Hrp Pathway of *Pseudomonas solanacearum*," EMBO J. 13:543-533 (1994), which is hereby incorporated by reference.

The hypersensitive response elicitor
 10 polypeptide or protein from *Xanthomonas campestris* pv. glycines has an amino acid sequence corresponding to SEQ. ID. No. 9 as follows:

15	Thr	Leu	Ile	Glu	Leu	Met	Ile	Val	Val	Ala	Ile	Ile	Ala	Ile	Leu	Ala
	1				5					10					15	
	Ala	Ile	Ala	Leu	Pro	Ala	Tyr	Gln	Asp	Tyr						
				20					25							

20 This sequence is an amino terminal sequence having 26 residues only from the hypersensitive response elicitor polypeptide or protein of *Xanthomonas campestris* pv. glycines. It matches with fimbrial subunit proteins
 25 determined in other *Xanthomonas campestris* pathovars.

The hypersensitive response elicitor polypeptide or protein from *Xanthomonas campestris* pv. *pelargonii* is heat stable, protease sensitive, and has a molecular weight of 20 kDa. It includes an amino acid
 30 sequence corresponding to SEQ. ID. No. 10 as follows:

35	Ser	Ser	Gln	Gln	Ser	Pro	Ser	Ala	Gly	Ser	Glu	Gln	Gln	Leu	Asp	Gln
	1				5					10					15	
	Leu	Leu	Ala	Met												
				20												

40 Isolation of *Erwinia carotovora* hypersensitive response elicitor protein or polypeptide is described in Cui et al., "The RsmA Mutants of *Erwinia carotovora*

subsp. *carotovora* Strain Ecc71 Overexpress *hrp* N_{Ecc} and Elicit a Hypersensitive Reaction-like Response in Tobacco Leaves," MPMI, 9(7):565-73 (1996), which is hereby incorporated by reference. The hypersensitive response elicitor proptein or polypeptide is shown in Ahmad et al., "Harpin is Not Necessary for the Pathogenicity of *Erwinia stewartii* on Maize," 8th Int'l. Cong. Molec. Plant-Microbe Interact., July 14-19, 1996 and Ahmad, et al., "Harpin is Not Necessary for the Pathogenicity of *Erwinia stewartii* on Maize," Ann. Mtg. Am. Phytopath. Soc., July 27-31, 1996, which are hereby incorporated by reference.

Hypersensitive response elicitor proteins or polypeptides from *Phytophthora parasitica*, *Phytophthora cryptogea*, *Phytophthora cinnamoni*, *Phytophthora capsici*, *Phytophthora megasperma*, and *Phytophthora citrophthora* are described in Kaman, et al., "Extracellular Protein Elicitors from *Phytophthora*: Most Specificity and Induction of Resistance to Bacterial and Fungal Phytopathogens," Molec. Plant-Microbe Interact., 6(1):15-25 (1993), Ricci et al., "Structure and Activity of Proteins from Pathogenic Fungi *Phytophthora* Eliciting Necrosis and Acquired Resistance in Tobacco," Eur. J. Biochem., 183:555-63 (1989), Ricci et al., "Differential Production of Parasiticein, and Elicitor of Necrosis and Resistance in Tobacco, by Isolates of *Phytophthora parasitica*," Plant Path. 41:298-307 (1992), Baillreul et al, "A New Elicitor of the Hypersensitive Response in Tobacco: A Fungal Glycoprotein Elicits Cell Death, Expression of Defence Genes, Production of Salicylic Acid, and Induction of Systemic Acquired Resistance," Plant J., 8(4):551-60 (1995), and Bonnet et al., "Acquired Resistance Triggered by Elicitors in Tobacco and Other Plants," Eur. J. Plant Path., 102:181-92 (1996), which are hereby incorporated by reference.

The above elicitors are exemplary. Other elicitors can be identified by growing fungi or bacteria that elicit a hypersensitive response under which genes encoding an elicitor are expressed. Cell-free
5 preparations from culture supernatants can be tested for elicitor activity (i.e. local necrosis) by using them to infiltrate appropriate plant tissues.

It is also possible to use fragments of the above hypersensitive response elicitor polypeptides or
10 proteins as well as fragments of full length elicitors from other pathogens, in the method of the present invention.

Suitable fragments can be produced by several means. In the first, subclones of the gene encoding a known elicitor protein are produced by conventional molecular genetic manipulation by subcloning gene
5 fragments. The subclones then are expressed *in vitro* or *in vivo* in bacterial cells to yield a smaller protein or a peptide that can be tested for elicitor activity according to the procedure described below.

As an alternative, fragments of an elicitor
10 protein can be produced by digestion of a full-length elicitor protein with proteolytic enzymes like chymotrypsin or *Staphylococcus* proteinase A, or trypsin. Different proteolytic enzymes are likely to cleave elicitor proteins at different sites based on the amino
15 acid sequence of the elicitor protein. Some of the fragments that result from proteolysis may be active elicitors of resistance.

In another approach, based on knowledge of the primary structure of the protein, fragments of the
20 elicitor protein gene may be synthesized by using the PCR technique together with specific sets of primers chosen to represent particular portions of the protein. These then would be cloned into an appropriate vector for

increase and expression of a truncated peptide or protein.

Chemical synthesis can also be used to make suitable fragments. Such a synthesis is carried out using known amino acid sequences for the elicitor being produced. Alternatively, subjecting a full length elicitor to high temperatures and pressures will produce fragments. These fragments can then be separated by conventional procedures (e.g., chromatography, SDS-PAGE).

An example of a useful fragment is the popA1 fragment of the hypersensitive response elicitor polypeptide or protein from *Pseudomonas solanacearum*. See Arlat, M., F. Van Gijsegem, J.C. Huet, J.C. Pemollet, and C.A. Boucher, "PopA1, a Protein Which Induces a Hypersensitive-like Response in Specific Petunia Genotypes is Secreted via the Hrp Pathway of *Pseudomonas solanacearum*," EMBO J. 13:543-53 (1994), which is hereby incorporated by reference. As to *Erwinia amylovora*, a suitable fragment can be, for example, either or both the polypeptide extending between and including amino acids 1 and 98 of SEQ. ID. No. 3 and the polypeptide extending between and including amino acids 137 and 204 of SEQ. ID. No. 3.

Variants may also (or alternatively) be modified by, for example, the deletion or addition of amino acids that have minimal influence on the properties, secondary structure and hydrophobic nature of the polypeptide. For example, a polypeptide may be conjugated to a signal (or leader) sequence at the N-terminal end of the protein which co-translationally or post-translationally directs transfer of the protein. The polypeptide may also be conjugated to a linker or other sequence for ease of synthesis, purification or identification of the polypeptide.

The protein or polypeptide of the present invention is preferably produced in purified form (preferably at least about 60%, more preferably 80%, pure) by conventional techniques. Typically, the protein or polypeptide of the present invention is produced but not secreted into the growth medium of recombinant host cells. Alternatively, the protein or polypeptide of the present invention is secreted into growth medium. In the case of unsecreted protein, to isolate the protein, the host cell (e.g., *E. coli*) carrying a recombinant plasmid is propagated, lysed by sonication, heat, or chemical treatment, and the homogenate is centrifuged to remove bacterial debris. The supernatant is then subjected to heat treatment and the hypersensitive response elicitor protein is separated by centrifugation. The supernatant fraction containing the polypeptide or protein of the present invention is subjected to gel filtration in an appropriately sized dextran or polyacrylamide column to separate the proteins. If necessary, the protein fraction may be further purified by ion exchange or HPLC.

The DNA molecule encoding the hypersensitive response elicitor polypeptide or protein can be incorporated in cells using conventional recombinant DNA technology. Generally, this involves inserting the DNA molecule into an expression system to which the DNA molecule is heterologous (i.e. not normally present). The heterologous DNA molecule is inserted into the expression system or vector in proper sense orientation and correct reading frame. The vector contains the necessary elements for the transcription and translation of the inserted protein-coding sequences.

U.S. Patent No. 4,237,224 to Cohen and Boyer, which is hereby incorporated by reference, describes the production of expression systems in the form of recombinant plasmids using restriction enzyme cleavage

and ligation with DNA ligase. These recombinant plasmids are then introduced by means of transformation and replicated in unicellular cultures including procaryotic organisms and eucaryotic cells grown in tissue culture.

5 Recombinant genes may also be introduced into viruses, such as vaccina virus. Recombinant viruses can be generated by transfection of plasmids into cells infected with virus.

10 Suitable vectors include, but are not limited to, the following viral vectors such as lambda vector system gt11, gt WES.tB, Charon 4, and plasmid vectors such as pBR322, pBR325, pACYC177, pACYC1084, pUC8, pUC9, pUC18, pUC19, pLG339, pR290, pKC37, pKC101, SV 40, pBluescript II SK +/- or KS +/- (see "Stratagene Cloning
15 Systems" Catalog (1993) from Stratagene, La Jolla, Calif, which is hereby incorporated by reference), pQE, pIH821, pGEX, pET series (see F.W. Studier et. al., "Use of T7 RNA Polymerase to Direct Expression of Cloned Genes," Gene Expression Technology vol. 185 (1990), which is
20 hereby incorporated by reference), and any derivatives thereof. Recombinant molecules can be introduced into cells via transformation, particularly transduction, conjugation, mobilization, or electroporation. The DNA sequences are cloned into the vector using standard
25 cloning procedures in the art, as described by Sambrook et al., Molecular Cloning: A Laboratory Manual, Cold Springs Laboratory, Cold Springs Harbor, New York (1989), which is hereby incorporated by reference.

30 A variety of host-vector systems may be utilized to express the protein-encoding sequence(s). Primarily, the vector system must be compatible with the host cell used. Host-vector systems include but are not limited to the following: bacteria transformed with bacteriophage DNA, plasmid DNA, or cosmid DNA;
35 microorganisms such as yeast containing yeast vectors;

mammalian cell systems infected with virus (e.g., vaccinia virus, adenovirus, etc.); insect cell systems infected with virus (e.g., baculovirus); and plant cells infected by bacteria. The expression elements of these
5 vectors vary in their strength and specificities. Depending upon the host-vector system utilized, any one of a number of suitable transcription and translation elements can be used.

Different genetic signals and processing events
10 control many levels of gene expression (e.g., DNA transcription and messenger RNA (mRNA) translation).

Transcription of DNA is dependent upon the presence of a promotor which is a DNA sequence that directs the binding of RNA polymerase and thereby
15 promotes mRNA synthesis. The DNA sequences of eucaryotic promoters differ from those of procaryotic promoters. Furthermore, eucaryotic promoters and accompanying genetic signals may not be recognized in or may not function in a procaryotic system, and, further,
20 procaryotic promoters are not recognized and do not function in eucaryotic cells.

Similarly, translation of mRNA in procaryotes depends upon the presence of the proper procaryotic signals which differ from those of eucaryotes. Efficient
25 translation of mRNA in procaryotes requires a ribosome binding site called the Shine-Dalgarno ("SD") sequence on the mRNA. This sequence is a short nucleotide sequence of mRNA that is located before the start codon, usually AUG, which encodes the amino-terminal methionine of the
30 protein. The SD sequences are complementary to the 3'-end of the 16S rRNA (ribosomal RNA) and probably promote binding of mRNA to ribosomes by duplexing with the rRNA to allow correct positioning of the ribosome. For a review on maximizing gene expression, see Roberts and

Lauer, Methods in Enzymology, 68:473 (1979), which is hereby incorporated by reference.

Promoters vary in their "strength" (i.e. their ability to promote transcription). For the purposes of
5 expressing a cloned gene, it is desirable to use strong promoters in order to obtain a high level of transcription and, hence, expression of the gene. Depending upon the host cell system utilized, any one of a number of suitable promoters may be used. For
10 instance, when cloning in *E. coli*, its bacteriophages, or plasmids, promoters such as the T7 phage promoter, *lac* promoter, *trp* promoter, *recA* promoter, ribosomal RNA promoter, the P_R and P_L promoters of coliphage lambda and others, including but not limited, to *lacUV5*, *ompF*, *bla*,
15 *lpp*, and the like, may be used to direct high levels of transcription of adjacent DNA segments. Additionally, a hybrid *trp-lacUV5* (*tac*) promoter or other *E. coli* promoters produced by recombinant DNA or other synthetic DNA techniques may be used to provide for transcription
20 of the inserted gene.

Bacterial host cell strains and expression vectors may be chosen which inhibit the action of the promoter unless specifically induced. In certain operations, the addition of specific inducers is
25 necessary for efficient transcription of the inserted DNA. For example, the *lac* operon is induced by the addition of lactose or IPTG (isopropylthio-beta-D-galactoside). A variety of other operons, such as *trp*, *pro*, etc., are under different controls.

30 Specific initiation signals are also required for efficient gene transcription and translation in procaryotic cells. These transcription and translation initiation signals may vary in "strength" as measured by the quantity of gene specific messenger RNA and protein
35 synthesized, respectively. The DNA expression vector,

which contains a promotor, may also contain any combination of various "strong" transcription and/or translation initiation signals. For instance, efficient translation in *E. coli* requires an SD sequence about 7-9 bases 5' to the initiation codon (ATG) to provide a ribosome binding site. Thus, any SD-ATG combination that can be utilized by host cell ribosomes may be employed. Such combinations include but are not limited to the SD-ATG combination from the *cro* gene or the *N* gene of coliphage lambda, or from the *E. coli* tryptophan E, D, C, B or A genes. Additionally, any SD-ATG combination produced by recombinant DNA or other techniques involving incorporation of synthetic nucleotides may be used.

Once the isolated DNA molecule encoding the hypersensitive response elicitor polypeptide or protein has been cloned into an expression system, it is ready to be incorporated into a host cell. Such incorporation can be carried out by the various forms of transformation noted above, depending upon the vector/host cell system. Suitable host cells include, but are not limited to, bacteria, virus, yeast, mammalian cells, insect, plant, and the like.

The method of the present invention can be utilized to treat a wide variety of plants or their seeds to enhance growth. Suitable plants include dicots and monocots. More particularly, useful crop plants can include: rice, wheat, barley, rye, cotton, sunflower, peanut, corn, potato, sweet potato, bean, pea, chicory, lettuce, endive, cabbage, cauliflower, broccoli, turnip, radish, spinach, onion, garlic, eggplant, pepper, celery, carrot, squash, pumpkin, zucchini, cucumber, apple, pear, melon, strawberry, grape, raspberry, pineapple, soybean, tobacco, tomato, sorghum, and sugarcane. Examples of suitable ornamental plants are: rose, *Saintpaulia*,

petunia, pelargonium, poinsettia, chrysanthemum, carnation, and zinnia.

The method of the present invention involving application of the hypersensitive response elicitor polypeptide or protein can be carried out through a variety of procedures when all or part of the plant is treated, including leaves, stems, roots, etc. This may (but need not) involve infiltration of the hypersensitive response elicitor polypeptide or protein into the plant. Suitable application methods include topical application (e.g., high or low pressure spraying), injection, dusting, and leaf abrasion proximate to when elicitor application takes place. When treating plant seeds, in accordance with the application embodiment of the present invention, the hypersensitive response elicitor protein or polypeptide can be applied by topical application (low or high pressure spraying), coating, immersion, dusting, or injection. Other suitable application procedures can be envisioned by those skilled in the art provided they are able to effect contact of the hypersensitive response elicitor polypeptide or protein with cells of the plant or plant seed. Once treated with the hypersensitive response elicitor of the present invention, the seeds can be planted in natural or artificial soil and cultivated using conventional procedures to produce plants. After plants have been propagated from seeds treated in accordance with the present invention, the plants may be treated with one or more applications of the hypersensitive response elicitor protein or polypeptide to enhance growth in the plants. Such propagated plants may, in turn, be useful in producing seeds or propagules (e.g., cuttings) that produce plants capable of enhanced growth.

The hypersensitive response elicitor polypeptide or protein can be applied to plants or plant

seeds in accordance with the present invention alone or in a mixture with other materials. Alternatively, the hypersensitive response elicitor polypeptide or protein can be applied separately to plants with other materials
5 being applied at different times.

A composition suitable for treating plants or plant seeds in accordance with the application embodiment of the present invention contains a hypersensitive response elicitor polypeptide or protein in a carrier.
10 Suitable carriers include water, aqueous solutions, slurries, or dry powders. In this embodiment, the composition contains greater than 0.5 nM hypersensitive response elicitor polypeptide or protein.

Although not required, this composition may
15 contain additional additives including fertilizer, insecticide, fungicide, nematocide, herbicide, and mixtures thereof. Suitable fertilizers include $(\text{NH}_4)_2\text{NO}_3$. An example of a suitable insecticide is Malathion. Useful fungicides include Captan.

20 Other suitable additives include buffering agents, wetting agents, coating agents, and abrading agents. These materials can be used to facilitate the process of the present invention. In addition, the hypersensitive response elicitor polypeptide or protein
25 can be applied to plant seeds with other conventional seed formulation and treatment materials, including clays and polysaccharides.

In the alternative embodiment of the present invention involving the use of transgenic plants and
30 transgenic seeds, a hypersensitive response elicitor polypeptide or protein need not be applied topically to the plants or seeds. Instead, transgenic plants transformed with a DNA molecule encoding a hypersensitive response elicitor polypeptide or protein are produced
35 according to procedures well known in the art, such as by

biolistics or *Agrobacterium* mediated transformation. Examples of suitable hypersensitive response elicitor polypeptides or proteins and the nucleic acid sequences for their encoding DNA are disclosed *supra*. Once
5 transgenic plants of this type are produced, the plants themselves can be cultivated in accordance with conventional procedure with the presence of the gene encoding the hypersensitive response elicitor resulting in enhanced growth of the plant. Alternatively,
10 transgenic seeds are recovered from the transgenic plants. These seeds can then be planted in the soil and cultivated using conventional procedures to produce transgenic plants. The transgenic plants are propagated from the planted transgenic seeds under conditions
15 effective to impart enhanced growth. While not wishing to be bound by theory, such growth enhancement may be RNA mediated or may result from expression of the elicitor polypeptide or protein.

When transgenic plants and plant seeds are used
20 in accordance with the present invention, they additionally can be treated with the same materials as are used to treat the plants and seeds to which a hypersensitive response elicitor polypeptide or protein is applied. These other materials, including
25 hypersensitive response elicitors, can be applied to the transgenic plants and plant seeds by the above-noted procedures, including high or low pressure spraying, injection, coating, dusting, and immersion. Similarly, after plants have been propagated from the transgenic
30 plant seeds, the plants may be treated with one or more applications of the hypersensitive response elicitor to enhance plant growth. Such plants may also be treated with conventional plant treatment agents (e.g., insecticides, fertilizers, etc.). The transgenic plants
35 of the present invention are useful in producing seeds or

propagules (e.g., cuttings) from which plants capable of enhanced growth would be produced.

EXAMPLES

5 **Example 1 - Effect of Treating Tomato Seeds with
 Erwinia amylovora Hypersensitive Response
 Elicitor on Germination Percentage**

10 Seeds of the Marglobe Tomato Variety were
 submerged in 40ml of *Erwinia amylovora* hypersensitive
 response elicitor solution ("harpin"). Harpin was
 prepared by growing *E. coli* strain DH5 containing the
 plasmid pCPP2139 (see Figure 1), lysing the cells by
15 sonication, heat treating by holding in boiling water for
 5 minutes before centrifuging to remove cellular debris,
 and precipitating proteins and other heat-labile
 components. The resulting preparation ("CFEP") was
 diluted serially. These dilutions (1:40, 1:80, 1:160,
20 1:320 and 1:640) contained 20, 10, 5, 2.5, and 1.25
 $\mu\text{gm/ml}$, respectively, of harpin based on Western Blot
 assay. Seeds were soaked in harpin or buffer in beakers
 on day 0 for 24 hours at 28°C in a growth chamber. After
 soaking, the seeds were sown in germination pots with
25 artificial soil on day 1. This procedure was carried out
 on 100 seeds per treatment.

Treatments:

- 30 1. Seeds in harpin (1:40) (20 $\mu\text{gm/ml}$).
 2. Seeds in harpin (1:80) (10 $\mu\text{gm/ml}$).
 3. Seeds in harpin (1:160) (5 $\mu\text{gm/ml}$).
 4. Seeds in harpin (1:320) (2.5 $\mu\text{gm/ml}$).
 5. Seeds in harpin (1:640) (1.25 $\mu\text{gm/ml}$).
 6. Seeds in buffer (5mM KPO_4 , pH 6.8).

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Table 1 - Number of Seedlings After Seed Treatment

Treatment	Number of seeds germinated				
	Day 0	Day 1	Day 5	Day 7	Day 9
5 Harpin seed soak (20 $\mu\text{gm/ml}$)	sowing	43	57	59	
Harpin seed soak (10 $\mu\text{gm/ml}$)	sowing	43	52	52	
Harpin seed soak (5 $\mu\text{gm/ml}$)	sowing	40	47	51	
Harpin seed soak (2.5 $\mu\text{gm/ml}$)	sowing	43	56	58	
10 Harpin seed soak (1.25 $\mu\text{gm/ml}$)	sowing	38	53	57	
Buffer seed soak	sowing	27	37	40	

As shown in Table 1, the treatment of tomato seeds with *Erwinia amylovora* hypersensitive response elicitor reduced the time needed for germination and greatly increased the percentage of germination.

Example 2 - Effect of Treating Tomato Seeds with *Erwinia amylovora* Hypersensitive Response Elicitor on Tomato Plant Height

Seeds of the Marglobe Tomato Variety were submerged in *Erwinia amylovora* harpin (1:15, 1:30, 1:60, and 1:120) or buffer in beakers on day 0 for 24 hours at 28°C in a growth chamber. After soaking, the seeds were sown in germination pots with artificial soil on day 1.

Ten uniform appearing plants per treatment were chosen randomly and measured. The seedlings were measured by ruler from the surface of soil to the top of plant.

Treatments:

1. Harpin (1:15) (52 $\mu\text{gm/ml}$).
2. Harpin (1:30) (26 $\mu\text{gm/ml}$).
3. Harpin (1:60) (13 $\mu\text{gm/ml}$).
4. Harpin (1:120) (6.5 $\mu\text{gm/ml}$).
5. Buffer (5mM KPO_4 , pH 6.8).

Table 2 - Seedling Height (cm) 15 Days After Seed Treatment.

Treat	Plants	1	2	3	4	5	6	7	8	9	10	Mean
52 μ gm/ml	10	5.6	5.8	5.8	5.6	6.4	6.0	5.8	5.4	5.8	5.6	5.7
26 μ gm/ml	10	6.8	7.2	6.6	7.0	6.8	5.8	7.0	7.4	7.2	7.2	7.0
13 μ gm/ml	10	5.8	5.6	6.0	5.6	6.0	5.8	5.6	5.8	6.0	5.6	5.9
6.5 μ gm/ml	10	5.8	5.2	6.6	5.4	5.2	5.6	5.6	5.6	5.4	5.8	5.4
Buffer	10	5.6	5.4	5.2	5.2	6.4	5.2	5.0	5.2	6.4	5.6	5.3

Table 3 - Seedling Height (cm) 21 Days After Seed Treatment.

Treat	Plants	1	2	3	4	5	6	7	8	9	10	Mean
52 μ gm/ml	10	7.6	7.8	7.6	7.6	7.8	7.8	7.8	7.4	7.6	7.6	7.7
26 μ gm/ml	10	8.2	8.2	9.0	9.0	8.4	8.6	8.6	9.0	9.2	9.0	8.6
13 μ gm/ml	10	6.6	6.6	6.8	6.8	6.8	6.8	6.6	7.2	7.0	7.2	6.9
6.5 μ gm/ml	10	6.8	6.6	6.6	6.4	6.8	6.6	6.6	6.6	6.6	6.8	6.7
Buffer	10	6.6	6.4	6.2	5.6	6.4	6.6	6.8	6.4	6.4	6.6	6.5

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Table 4 - Seedling Height (cm) 27 Days After Seed Treatment.

Treat	1	2	3	4	5	6	7	8	9	10	Mean
52 μ gm/ml	10.2	10.6	10.4	10.6	10.4	10.6	10.8	10.4	10.8	10.6	10.5
26 μ gm/ml	11.6	11.4	11.6	11.8	11.8	11.8	11.6	11.4	11.6	11.4	11.6
13 μ gm/ml	9.8	9.6	9.8	9.6	9.8	9.8	9.6	9.4	9.6	9.8	9.7
6.5 μ gm/ml	9.4	9.4	9.6	9.4	9.6	9.4	9.6	9.6	9.4	9.2	9.5
Buffer	9.6	10.2	10.0	9.8	10.0	10.2	10.0	10.0	10.4	9.6	10.0

Table 5 - Summary--Mean Height of Tomato Plants after Treatment.

5	Treatment	Mean height of tomato plants(cm)				
	Day 0	Day 1	Day 15	Day 21	Day 27	
10	Harpin seed soak (1:15)	sowing	5.7	7.7	10.5	
	Harpin seed soak (1:30)	sowing	7.0	8.6	11.6	
	Harpin seed soak (1:60)	sowing	5.9	6.9	9.7	
	Harpin seed soak (1:120)	sowing	5.4	6.7	9.5	
	Buffer seed soak	sowing	5.3	6.5	10.0	

As shown in Tables 2-5, the treatment of tomato seeds with *Erwinia amylovora* hypersensitive response elicitor increased plant growth. A 1:30 dilution had the greatest effect -- a 16% increase in seedling height.

Example 3 - Effect of Treating Tomato Plants with *Erwinia amylovora* Hypersensitive Response Elicitor on Tomato Plant Height

When *Marglobe* tomato plants were 4 weeks old, they were sprayed with 6 ml/plant of *Erwinia amylovora* harpin solution containing 13 $\mu\text{gm/ml}$ (1:60) or 8.7 $\mu\text{gm/ml}$ (1:90) of harpin or buffer (5mM KPO_4) in a growth chamber at 28°C. The heights of tomato plants were measured 2 weeks after spraying harpin (6-week-old tomato plants) and 2 weeks plus 5 days after spraying. Ten uniform appearing plants per treatment were chosen randomly and measured. The seedlings were measured by ruler from the surface of soil to the top of plant.

Treatments:

1. Harpin (1:60) (13 $\mu\text{gm/ml}$).
2. Harpin (1:90) (8.7 $\mu\text{gm/ml}$).
3. Buffer (5mM KPO_4 , pH 6.8).

Table 6 - Mean Height of Tomato Plants after Treatment With Harpin.

5	Operation and Treatment			Mean height (cm) of tomato plants	
	Day 0	Day 14	Day 28	Day 42	Day 47
10	sowing	transplant	harpin 1:60 (13 µgm/ml)	35.5	36.0
	sowing	transplant	harpin 1:90 (8.7 µgm/ml)	35.7	36.5
15	sowing	transplant	buffer	32.5	33.0

As shown in Table 6, spraying tomato seedlings with *Erwinia amylovora* hypersensitive response elicitor can increase growth of tomato plants. Similar increases in growth were noted for the two doses of the hypersensitive response elicitor tested compared with the buffer-treated control.

Example 4 - Effect of Treating Tomato Seeds with *Erwinia amylovora* Hypersensitive Response Elicitor on Tomato Plant Height

Marglobe tomato seeds were submerged in *Erwinia amylovora* hypersensitive response elicitor solution ("harpin") (1:40, 1:80, 1:160, 1:320, and 1:640) or buffer in beakers on day 0 for 24 hours at 28°C in the growth chamber. After soaking seeds in harpin or buffer, they were sown in germination pots with artificial soil on day 1. Ten uniform appearing plants per treatment were chosen randomly and measured. The seedlings were measured by ruler from the surface of soil to the top of plant.

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Table 7 - Seedling Height (cm) 12 Days After Seed Treatment.

Treat	Plants	1	2	3	4	5	6	7	8	9	10	Mean
20 μ gm/ml	10	6.5	6.8	6.8	6.5	6.4	6.0	6.8	6.4	6.8	6.6	6.6
10 μ gm/ml	10	6.8	6.2	6.9	6.8	6.8	6.8	6.6	6.8	6.8	6.4	6.6
5 μ gm/ml	10	6.2	6.4	6.0	6.6	6.4	6.2	6.6	6.2	6.0	6.6	6.3
2.5 μ gm/ml	10	6.4	6.2	6.6	6.0	6.2	6.4	6.0	6.0	6.2	6.2	6.2
1.25 μ gm/ml	10	6.2	6.2	6.0	6.4	6.0	6.0	6.0	6.2	6.4	6.2	6.2
Buffer	10	5.8	6.0	6.2	6.2	5.8	6.8	6.0	6.2	6.0	6.0	6.0

Table 8 - Seedling Height (cm) 14 Days After Seed Treatment.

Treat	Plants	1	2	3	4	5	6	7	8	9	10	Mean
20 μ gm/ml	10	7.8	7.8	8.2	8.0	8.2	8.4	7.8	8.4	7.6	7.8	8.0
10 μ gm/ml	10	8.6	8.8	8.4	9.2	8.4	8.6	7.8	7.8	8.4	8.4	8.4
5 μ gm/ml	10	9.8	8.2	9.8	9.6	9.2	9.4	8.6	9.2	9.0	8.6	9.2
2.5 μ gm/ml	10	8.8	8.6	8.6	8.8	7.8	8.6	8.4	9.0	9.0	7.8	8.8
1.25 μ gm/ml	10	8.4	7.8	8.8	9.0	8.6	8.4	8.0	8.2	8.4	8.2	8.2
Buffer	10	7.2	8.2	7.4	7.6	7.8	7.6	7.8	7.8	7.8	7.6	7.6

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Table 9 - Seedling Height (cm) 17 Days After Seed Treatment.

Treat	Plants	1	2	3	4	5	6	7	8	9	10	Mean
20 μ gm/ml	10	11.2	11.6	11.4	11.6	11.4	11.2	11.8	11.4	11.8	11.6	11.5
10 μ gm/ml	10	13.4	13.4	13.8	13.2	13.4	12.6	12.4	13.4	13.2	13.4	13.2
5 μ gm/ml	10	13.6	12.4	13.6	13.2	14.2	13.8	12.6	13.4	13.8	13.6	13.5
2.5 μ gm/ml	10	11.6	12.4	12.4	11.8	11.6	12.2	12.6	11.8	12.0	11.6	12.0
1.25 μ gm/ml	10	12.8	12.6	12.0	12.4	11.6	11.8	12.2	11.4	11.2	11.4	11.9
Buffer	10	10.0	10.4	10.6	10.6	10.4	10.4	10.8	10.2	10.4	10.0	10.4

Table 10 -Summary - Mean Height of Tomato Plants After Treatment

Operation and Treatment	Mean height of tomato plants (cm)				
	Day 0	Day 1	Day 12	Day 14	Day 17
Harpin seed soak (20 μ gm/ml)		sowing	6.6	8.0	11.5
Harpin seed soak (10 μ gm/ml)		sowing	6.6	8.4	13.2
Harpin seed soak (5 μ gm/ml)		sowing	6.3	9.2	13.5
Harpin seed soak (2.5 μ gm/ml)		sowing	6.2	8.4	12.0
Harpin seed soak (1.25 μ gm/ml)		sowing	6.2	8.2	11.9
Buffer seed soak		sowing	6.0	7.6	10.4

As shown in Tables 7-10, the treatment of tomato seeds with *Erwinia amylovora* hypersensitive response elicitor can increase growth of tomato plants. A 1:160 dilution (5 $\mu\text{g}/\text{ml}$ harpin) had the greatest effect -- seedling height was increased more than 20% over the buffer treated plants.

Example 5 - Effect of Treating Tomato Seeds with *Erwinia amylovora* Hypersensitive Response Elicitor on Seed Germination Percentage

Marglobe tomato seeds were submerged in 40ml of *Erwinia amylovora* hypersensitive response elicitor ("harpin") solution (dilutions of CFEP from *E. coli* DH5 (pCPP2139) of 1:50 or 1:100 which contained, respectively, 8 $\mu\text{gm}/\text{ml}$ and 4 $\mu\text{gm}/\text{ml}$ of hypersensitive response elicitor) and buffer in beakers on day 0 for 24 hours at 28°C in a growth chamber. After soaking, the seeds were sown in germination pots with artificial soil on day 1. This treatment was carried out on 20 seeds per pot and 4 pots per treatment.

Treatments:

1. Harpin (8 $\mu\text{gm}/\text{ml}$).
2. Harpin (8 $\mu\text{gm}/\text{ml}$).
3. Harpin (8 $\mu\text{gm}/\text{ml}$).
4. Harpin (8 $\mu\text{gm}/\text{ml}$).
5. Harpin (4 $\mu\text{gm}/\text{ml}$).
6. Harpin (4 $\mu\text{gm}/\text{ml}$).
7. Harpin (4 $\mu\text{gm}/\text{ml}$).
8. Harpin (4 $\mu\text{gm}/\text{ml}$).
9. Buffer (5mM KPO_4 , pH 6.8).
10. Buffer (5mM KPO_4 , pH 6.8).
11. Buffer (5mM KPO_4 , pH 6.8).
12. Buffer (5mM KPO_4 , pH 6.8).

Table 11 - Number of Seedlings After Seed Treatment With Harpin

5	Operation and Treatment		Number of seeds germinated (out of a total of 20)					
			Day 0	Day 1	Day 5	Day 42	Day 47	
					Mean	Mean	Mean	
10	Harpin (8 µgm/ml)	sowing		11		15	19	
	Harpin (8 µgm/ml)	sowing		13		17	20	
	Harpin (8 µgm/ml)	sowing		10		13	16	
	Harpin (8 µgm/ml)	sowing		9	10.8	15	15.0	17.8
15	Harpin (4 µgm/ml)	sowing		11		17	17	
	Harpin (4 µgm/ml)	sowing		15		17	18	
	Harpin (4 µgm/ml)	sowing		9		12	14	
	Harpin (4 µgm/ml)	sowing		9	11.0	14	15.0	16.3
20	Buffer	sowing		11		11	14	
	Buffer	sowing		9		14	15	
	Buffer	sowing		10		14	14	
	Buffer	sowing		10	10.0	12	12.8	14.3

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As shown in Table 11, treatment of tomato seeds with *Erwinia amylovora* hypersensitive response elicitor can increase germination rate and level of tomato seeds. The higher dose used appeared to be more effective than buffer at the end of the experiment.

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Example 6 - Effect on Plant Growth of Treating Tomato Seeds with Proteins Prepared from *E. coli* Containing a Hypersensitive Response Elicitor Encoding Construct, pCPP2139, or Plasmid Vector pCPP50

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Marglobe tomato seeds were submerged in *Erwinia amylovora* hypersensitive response elicitor ("harpin") (from *E. coli* DH5α(pCPP2139) (Figure 1) or vector preparation (from DH5α(pCPP50) (Figure 2) with added BSA protein as control. The control vector preparation contained, per ml, 33.6 µl of BSA (10 mg/ml) to provide about the same amount of protein as contained in the

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pCPP2139 preparation due to harpin. Dilutions of 1:50 (8.0 $\mu\text{g/ml}$), 1:100 (4.0 $\mu\text{g/ml}$), and 1:200 (2.0 $\mu\text{g/ml}$) were prepared in beakers on day 1, and seed was submerged for 24 hours at 28°C in a controlled environment chamber.

- 5 After soaking, seeds were sown in germination pots with artificial soil on day 2. Ten uniform appearing plants per treatment were chosen randomly and measured at three times after transplanting. The seedlings were measured by ruler from the surface of soil to the top of plant.

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Treatments:

- | | | | | |
|----|----|--------------|-------|-------------------------|
| | 1. | Harpin | 1:50 | (8.0 $\mu\text{g/ml}$) |
| | 2. | Harpin | 1:100 | (4.0 $\mu\text{g/ml}$) |
| | 3. | Harpin | 1:200 | (2.0 $\mu\text{g/ml}$) |
| 15 | 4. | Vector + BSA | 1:50 | (0 harpin) |
| | 5. | Vector + BSA | 1:100 | (0 harpin) |
| | 6. | Vector + BSA | 1:200 | (0 harpin) |

Table 14 - Seedling Height (cm) 26 Days After Seed Treatment.

Treat.	Harpin	1	2	3	4	5	6	7	8	9	10	Mean
H1:50	8.0	7.6	8.4	8.8	6.8	9.6	8.2	7.4	9.8	9.2	9.0	8.5
H1:100	4.0	12.0	11.4	11.2	11.0	10.8	12.0	11.2	11.6	10.4	10.2	11.2
H1:200	2.0	10.6	11.2	11.6	10.2	11.0	10.8	10.0	11.8	10.2	10.6	10.8
V1:50	0	9.0	9.4	8.8	8.4	9.6	9.2	9.2	8.6	8.0	9.4	9.2
V1:100	0	9.2	10.0	9.8	9.6	8.4	9.4	9.6	9.8	8.0	9.6	9.3
V1:200	0	8.8	9.6	8.2	9.2	8.4	8.0	9.8	9.0	9.4	9.2	9.0

Table 15 - Mean Height of Tomato Plants After Treatment

Operation and Treatment		Mean height of tomato plants (cm)			
Day 1	Day 2	Day 18	Day 22	Day 26	
Harpin (1:50) (8.0 μ gm/ml)	sowing	4.7	5.2	8.5	
Harpin (1:100) (4.0 μ gm/ml)	sowing	5.8	7.2	11.2	
Harpin (1:200) (2.0 μ gm/ml)	sowing	5.1	7.0	10.8	
Vector + BSA (1:50) (0)	sowing	4.9	5.8	9.2	
Vector + BSA (1:100) (0)	sowing	4.8	5.7	9.3	
Vector + BSA (1:200) (0)	sowing	4.9	5.9	9.0	

As shown in Tables 12-15, treatment with *E. coli* containing the gene encoding the *Erwinia amylovora* hypersensitive response elicitor can increase growth of tomato plants. The 1:100 dilution (4.0 µg/ml) had the greatest effect, while higher and lower concentrations had less effect. Mean seedling height for treatment with 4.0 µg/ml of harpin was increased about 20% relative to vector control preparation, which contained a similar amount of non-harpin protein. Components of the lysed cell preparation from the strain *E. coli* DH5α(pCPP50); which harbors the vector of the *hrpN* gene in *E. coli* strain DH5α(pCPP2139), do not have the same growth-promoting effect as the harpin-containing preparation, even given that it is supplemented with BSA protein to the same extent as the DH5α(pCPP2139) preparation, which contains large amounts of harpin protein.

Example 7 - Effect on Tomato Plant Growth of Treating Tomato Seeds with Proteins Prepared from *E. coli* Containing a Hypersensitive Response Elicitor Encoding Construct, pCPP2139, or its Plasmid Vector pCPP50

Marglobe tomato seeds were submerged in *Erwinia amylovora* hypersensitive response elicitor solution ("harpin") (from the harpin encoding plasmid pCPP2139 vector) and from pCPP50 vector-containing solution at dilutions of 1:25, 1:50, and 1:100 in beakers on day 1 for 24 hours at 28°C in a growth chamber. After soaking seeds, they were sown in germination pots with artificial soil on day 2. Ten uniform appearing plants per treatment were chosen randomly and measured. The seedlings were measured by ruler from the surface of soil to the top of plant.

Table 1. Demographic characteristics of the study population	
Age (years)	Mean (SD)
18-24	20.5 (2.5)
25-34	29.5 (4.5)
35-44	39.5 (5.5)
45-54	49.5 (6.5)
55-64	59.5 (7.5)
65-74	69.5 (8.5)
75-84	79.5 (9.5)
85-94	89.5 (10.5)
95-104	99.5 (11.5)
105-114	109.5 (12.5)
115-124	119.5 (13.5)
125-134	129.5 (14.5)
135-144	139.5 (15.5)
145-154	149.5 (16.5)
155-164	159.5 (17.5)
165-174	169.5 (18.5)
175-184	179.5 (19.5)
185-194	189.5 (20.5)
195-204	199.5 (21.5)
205-214	209.5 (22.5)
215-224	219.5 (23.5)
225-234	229.5 (24.5)
235-244	239.5 (25.5)
245-254	249.5 (26.5)
255-264	259.5 (27.5)
265-274	269.5 (28.5)
275-284	279.5 (29.5)
285-294	289.5 (30.5)
295-304	299.5 (31.5)
305-314	309.5 (32.5)
315-324	319.5 (33.5)
325-334	329.5 (34.5)
335-344	339.5 (35.5)
345-354	349.5 (36.5)
355-364	359.5 (37.5)
365-374	369.5 (38.5)
375-384	379.5 (39.5)
385-394	389.5 (40.5)
395-404	399.5 (41.5)
405-414	409.5 (42.5)
415-424	419.5 (43.5)
425-434	429.5 (44.5)
435-444	439.5 (45.5)
445-454	449.5 (46.5)
455-464	459.5 (47.5)
465-474	469.5 (48.5)
475-484	479.5 (49.5)
485-494	489.5 (50.5)
495-504	499.5 (51.5)
505-514	509.5 (52.5)
515-524	519.5 (53.5)
525-534	529.5 (54.5)
535-544	539.5 (55.5)
545-554	549.5 (56.5)
555-564	559.5 (57.5)
565-574	569.5 (58.5)
575-584	579.5 (59.5)
585-594	589.5 (60.5)
595-604	599.5 (61.5)
605-614	609.5 (62.5)
615-624	619.5 (63.5)
625-634	629.5 (64.5)
635-644	639.5 (65.5)
645-654	649.5 (66.5)
655-664	659.5 (67.5)
665-674	669.5 (68.5)
675-684	679.5 (69.5)
685-694	689.5 (70.5)
695-704	699.5 (71.5)
705-714	709.5 (72.5)
715-724	719.5 (73.5)
725-734	729.5 (74.5)
735-744	739.5 (75.5)
745-754	749.5 (76.5)
755-764	759.5 (77.5)
765-774	769.5 (78.5)
775-784	779.5 (79.5)
785-794	789.5 (80.5)
795-804	799.5 (81.5)
805-814	809.5 (82.5)
815-824	819.5 (83.5)
825-834	829.5 (84.5)
835-844	839.5 (85.5)
845-854	849.5 (86.5)
855-864	859.5 (87.5)
865-874	869.5 (88.5)
875-884	879.5 (89.5)
885-894	889.5 (90.5)
895-904	899.5 (91.5)
905-914	909.5 (92.5)
915-924	919.5 (93.5)
925-934	929.5 (94.5)
935-944	939.5 (95.5)
945-954	949.5 (96.5)
955-964	959.5 (97.5)
965-974	969.5 (98.5)
975-984	979.5 (99.5)
985-994	989.5 (100.5)
995-1004	999.5 (101.5)
1005-1014	1009.5 (102.5)
1015-1024	1019.5 (103.5)
1025-1034	1029.5 (104.5)
1035-1044	1039.5 (105.5)
1045-1054	1049.5 (106.5)
1055-1064	1059.5 (107.5)
1065-1074	1069.5 (108.5)
1075-1084	1079.5 (109.5)
1085-1094	1089.5 (110.5)
1095-1104	1099.5 (111.5)
1105-1114	1109.5 (112.5)
1115-1124	1119.5 (113.5)
1125-1134	

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Table 18 - Mean Height of Tomato Plants After Treatment.

5	Operation and Treatment	Mean height of tomato plants (cm)			
		Day 1	Day 2	Day 11	Day 14
10	Harpin seed soak (16 μ gm/ml)	sowing	4.5	7.4	
	Harpin seed soak (8 μ gm/ml)	sowing	5.5	8.2	
	Harpin seed soak (4 μ gm/ml)	sowing	5.1	7.9	
	Vector seed soak (16 μ gm/ml)	sowing	4.5	6.7	
	Vector seed soak (8 μ gm/ml)	sowing	4.4	6.7	
	Vector seed soak (4 μ gm/ml)	sowing	4.3	6.4	
15					

As shown in Tables 16-18, treatment with *Erwinia amylovora* hypersensitive response elicitor can increase growth of tomato plants. A 1:50 dilution (8 μ g/ml hypersensitive response elicitor) had the greatest effect with seedling height being increased by about 20% over the control.

Example 8 - Effect of Cell-Free *Erwinia amylovora* Hypersensitive Response Elicitor on Growth of Potato

Three-week-old potato plants, variety *Norchip*, were grown from tuber pieces in individual containers. The foliage of each plant was sprayed with a solution containing *Erwinia amylovora* hypersensitive response elicitor ("harpin"), or a control solution containing proteins of *E. coli* and those of the vector pCPP50 ("vector"), diluted 1:50, 1:100, and 1:200. On day 20, 12 uniform appearing plants were chosen randomly for each of the following treatments. One plant from each treatment was maintained at 16°C, in a growth chamber, while two plants from each treatment were maintained on a greenhouse bench at 18-25°C. Twenty-five days after treatment, the shoots (stems) on all plants were measured individually.

Table 19 - Length of Potato Stems of Plants at 16°C

Treatment on day 20	Length of potato stems (cm) on day 45					
	stem 1	stem 2	stem 3	stem 4	stem 5	Plant Mean
Harpin 1:50	43.0	39.5	42.5	34.0	38.0	39.4
Harpin 1:100	42.0	38.5	(2 branch)			40.3
Harpin 1:200	35.5	30.5	31.5	(3 branch)		32.5
Vector 1:50	34.0	32.0	31.5	28.0	27.5 (5 branch)	30.6
Vector 1:100	30.0	33.5	33.0	30.0	28.0	31.3
Vector 1:200	33.5	31.5	32.5	(3 branch)		32.5

Table 20 - Length of Potato Stems of Plants on a Greenhouse Bench

Treatment on day 20	Length of potato stems (cm) on day 45						Treat. Mean
	stem 1	stem 2	stem 3	stem 4	stem 5	Plant	
Harpin 1:50	65.5	58.5	57.5	62.5	68.5	62.5	64.2
Harpin 1:100	62.5	67.0	65.0	69.0	(4 branch)	55.9	
Harpin 1:200	70.5	73.5	74.0	80.5	(4 branch)	74.6	
Vector 1:50	83.0	80.5	76.5	76.0	81.5	79.5	77.1
Vector 1:100	56.5	59.5	50.5	53.0	55.5	53.9	
Vector 1:200	57.0	59.5	69.5	(3 branch)		62.0	58.0
Vector 1:50	53.0	62.0	59.5	62.5	(4 branch)	59.3	
Vector 1:100	52.0	46.0	61.5	56.5	61.5	55.8	57.6
Vector 1:200	62.0	51.5	66.0	67.5	62.0	62.0	
Vector 1:50	61.5	62.5	59.0	65.5	63.0	62.5	62.3
Vector 1:100	62.0	66.0	(2 branch)			64.0	
Vector 1:200	61.0	60.0	63.5	(3 branch)		61.5	62.8

As shown in Tables 19 and 20, treatment of potato plants with *Erwinia amylovora* hypersensitive response elicitor enhanced shoot (stem) growth. Thus, overall growth, as judged by both the number and mean
5 lengths of stems, were greater in the harpin-treated plants in both the greenhouse and growth chamber-grown plants. The potato plants treated with the medium dose of harpin (8 μ g/ml) seemed enhanced in their stem growth more than those treated with either higher or lower
10 doses. Treatment with the medium dose of harpin resulted in greater growth under both growing conditions.

Example 9 - Effect of Spraying Tomatoes With a Cell-Free Elicitor Preparation Containing the *Erwinia amylovora* Harpin
15

Marglobe tomato plants were sprayed with harpin preparation (from *E. coli* DH5 α (pCPP2139)) or vector preparation (from *E. coli* DH5 α (pCPP50)) with added BSA
20 protein as control 8 days after transplanting. The control vector preparation contained, per ml, 33.6 μ l of BSA (10 mg/ml) to provide about the same amount of protein as contained in the pCPP2139 preparation due to harpin. Dilutions of 1:50 (8.0 μ g/ml), 1:100
25 (4.0 μ g/ml), and 1:200 (2.0 μ g/ml) were prepared and sprayed on the plants to runoff with an electricity-powered atomizer. Fifteen uniform appearing plants per treatment were chosen randomly and assigned to treatment. The plants were maintained at 28°C in a controlled
30 environment chamber before and after treatment.

Overall heights were measured several times after treatment from the surface of soil to the top of the plant. The tops of the tomato plants were weighed immediately after cutting the stems near the surface of
35 the soil.

Table 21 -Tomato plant height (cm) 1 day after spray treatment

Treat	1	2	3	4	5	6	7	8	9	10	11	12	13	14	15	Mean
H 50	5.4	5.0	5.6	5.0	5.2	4.8	5.0	5.2	5.4	5.0	5.6	4.8	4.6	5.0	5.8	5.16
H 100	5.0	5.2	5.0	5.4	5.0	5.0	5.2	4.8	5.6	5.2	5.4	5.0	4.8	5.0	5.2	5.15
H 200	5.0	4.6	5.4	4.6	5.0	5.2	5.4	4.8	5.0	5.2	5.4	5.2	5.0	5.2	5.0	5.13
V 50	5.2	4.6	4.8	5.0	5.6	4.8	5.6	5.2	5.6	5.4	5.2	5.8	5.0	4.8	5.2	5.15
V 100	5.2	4.8	5.2	5.6	5.6	4.8	5.4	5.2	5.0	4.8	5.0	4.8	5.6	5.2	5.4	5.13
V 200	5.2	5.4	5.0	5.4	5.2	5.4	5.0	5.2	5.4	5.2	4.6	4.8	5.2	5.0	5.4	5.16

Table 22 -Tomato plant height (cm) 15 days after spray treatment

Treat	1	2	3	4	5	6	7	8	9	10	11	12	13	14	15	Mean
H 50	22.0	21.0	22.0	21.5	23.0	22.0	23.5	25.0	22.0	20.5	21.0	23.5	22.0	22.5	21.0	22.2
H 100	26.0	26.5	27.0	25.0	27.5	26.0	28.0	29.0	28.5	26.0	27.5	28.0	28.0	29.0	26.0	27.5
H 200	24.5	26.0	25.0	26.0	26.5	27.5	28.5	28.0	26.0	24.0	26.5	24.5	26.0	24.0	27.5	26.0
V 50	23.5	21.5	20.5	22.5	20.5	21.0	22.0	23.5	22.0	20.5	22.0	21.0	20.5	22.5	21.5	21.7
V 100	22.5	21.0	20.5	23.0	22.0	20.0	20.5	20.0	21.0	22.0	23.0	20.0	22.0	21.0	22.5	21.4
V 200	21.5	20.5	23.5	20.5	22.0	22.0	22.5	20.0	22.0	23.5	23.5	22.0	20.0	23.0	21.0	21.8

Table 23 -Tomato plant height (cm) 21 days after spray treatment

Treat	1	2	3	4	5	6	7	8	9	10	11	12	13	14	15	Mean
H 50	28.5	28.0	27.5	26.0	27.0	28.5	28.5	29.0	30.0	28.5	29.0	27.0	28.5	28.0	27.0	28.1
H 100	37.0	38.0	37.5	39.0	37.0	38.5	36.0	38.0	37.0	38.5	37.0	36.0	37.0	37.0	38.5	37.5
H 200	34.5	34.0	36.0	33.5	32.0	34.5	32.5	34.0	32.0	36.5	30.5	32.0	30.0	32.5	34.0	33.2
V 50	30.0	28.0	28.0	28.5	30.0	27.0	26.5	28.0	29.5	28.5	29.5	28.5	27.0	29.5	28.5	28.3
V 100	28.0	27.5	30.0	29.5	28.5	29.0	30.0	26.5	27.5	28.0	30.0	29.0	28.5	28.0	29.5	28.6
V 200	28.5	30.5	27.0	29.0	28.5	27.5	29.0	30.0	28.0	28.5	29.0	30.5	27.5	28.5	28.0	28.7

Table 24 -Mean Height of Tomato Plants After Spraying

Treatment (Dil. & harpin)	Mean height of tomato plants (cm)			
	Days After Treatment			
	Day 1	Day 11	Day 14	
Harpin 1:50	(8.0 µg/ml)	5.16	22.2	28.1
Harpin 1:100	(4.0 µg/ml)	5.15	27.5	37.5
Harpin 1:200	(2.0 µg/ml)	5.13	26.0	33.2
Vector + BSA 1:50	(0)	5.15	21.7	28.5
Vector + BSA 1:100	(0)	5.13	21.4	28.6
Vector + BSA 1:200	(0)	5.16	21.8	28.7

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Table 25 - Fresh Weight of Tomato Plants (g/plant)
21 Days After Spray Treatment

Treat	1	2	3	4	5	6	7	8	9	10	11	12	13	14	15	Mean
H 50	65.4	60.3	58.9	73.2	63.8	70.1	58.4	60.1	62.7	55.6	58.3	68.9	58.2	64.2	56.4	62.3
H 100	84.3	68.8	74.6	66.7	78.5	58.9	76.4	78.6	84.8	78.4	86.4	66.5	76.5	82.4	80.5	76.2
H 200	80.1	76.5	68.4	79.5	64.8	79.6	76.4	80.2	66.8	72.5	78.8	72.3	62.8	76.4	73.2	73.9
V 50	64.0	56.8	69.4	72.3	56.7	66.8	71.2	62.3	61.0	62.5	63.4	58.3	72.1	67.8	67.0	64.7
V 100	62.8	58.4	70.2	64.2	58.1	72.7	68.4	53.6	67.5	66.3	59.3	68.2	71.2	65.2	59.2	64.4
V 200	64.2	59.6	70.2	66.6	64.3	60.4	60.8	56.7	71.8	60.6	63.6	58.9	68.3	57.2	60.0	62.9

A single spray of tomato seedlings with harpin, in general, resulted in greater subsequent growth than spray treatment with the control (vector) preparation, which had been supplemented with BSA protein. Enhanced growth in the harpin-treated plants was seen in both plant height and fresh weight measurements. Of the three concentrations tested, the two lower ones resulted in more plant growth (based on either measure) than the higher dose (8.0 $\mu\text{g/ml}$). There was little difference in the growth of plants treated with the two lower (2 and 4 $\mu\text{g/ml}$) concentrations. Components of the lysed cell preparation from the strain *E. coli* DH5 α (pCPP50), which harbors the vector of the *hrpN* gene in *E. coli* strain DH5 α (pCPP2139), do not have the same growth-promoting effect as the harpin-containing preparation, even though it is supplemented with BSA protein to the same extent as the DH5 α (pCPP2139) preparation, which contains large amounts of harpin protein. Thus, this experiment demonstrates that harpin is responsible for enhanced plant growth.

Example 10 - Early Coloration and Early Ripening of
Raspberry Small Fruits

A field trial was conducted to evaluate the effect of hypersensitive response elicitor ("harpin") treatment on yield and ripening parameters of raspberry cv. Canby. Established plants were treated with harpin at 2.5 mg/100 square feet in plots 40 feet long x 3 feet wide (1 plant wide), untreated ("Check"), or treated with the industry standard chemical Ronilan at recommended rates ("Ronilan"). Treatments were replicated four times and arranged by rep in an experimental field site. Treatments were made beginning at 5-10% bloom followed by two applications at 7-10 day intervals. The first two harvests were used to evaluate disease control and fruit

yield data was collected from the last two harvests. Observations indicated harpin-treated fruits were larger and exhibited more redness than untreated fruits, indicating ripening was accelerated by 1-2 weeks. The
 5 number of ripe fruits per cluster bearing a minimum of ten fruits was determined at this time and is summarized in Table 26. Harpin treated plots had more ripe fruits per 10-berry cluster than either the check or Ronilan treatments. Combined yields from the last two harvests
 10 indicated increased yield in harpin and Ronilan treated plots over the untreated control (Table 27).

Table 26 - Number of Ripe Raspberry Fruits Per Clusters With Ten Berries or More on June 20, 1996.

<u>Treatment</u>	<u>Ripe fruit/10 berry clusters</u>	<u>% of Control</u>
Check	2.75	100.0
Ronilan	2.75	100.0
Harpin	7.25	263.6

Table 27 - Mean Raspberry Fruit Yield by Weight (lbs.) Combined in Last Two Harvest.

<u>Treatment</u>	<u>Total Yield</u>	<u>% of Control</u>
Check	32.5	100.0
Ronilan	37.5	115.4
Harpin	39.5	121.5

Example 11 - Growth Enhancement For Snap Beans

Snap beans of the variety Bush Blue Lake were treated by various methods, planted in 25-cm-d plastic
 5 pots filled with commercial potting mix, and placed in an open greenhouse for the evaluation of growth parameters. Treatments included untreated bean seeds ("Check"), seeds treated with a slurry of 1.5% methyl cellulose prepared with water as diluent ("M/C"), seeds treated with 1.5%
 10 methyl cellulose followed by a foliar application of hypersensitive response elicitor ("harpin") at 0.125

mg/ml ("M/C+H"), and seeds treated with 1.5% methyl cellulose plus harpin spray dried at 5.0 µg harpin per 50 seeds followed by a foliar application of harpin at 0.125 mg/ml ("M/C-SD+H"). Seeds were sown on day 0, planted 3 per pot, and thinned to 1 plant per pot upon germination. Treatments were replicated 10 times and randomized by rep in an open greenhouse. Bean pods were harvested after 64 days, and fresh weights of bean pods of marketable size (>10 cm x 5 cm in size) were collected as yield. Data were analyzed by analysis of variance with Fisher's LSD used to separate treatment means.

Table 28 - Effect of *Erwinia amylovora* Harpin Treatment by Various Methods on Yield of Market Sized Snap Bean Pods

	<u>Treatment</u>	<u>Marketable Yield, g¹</u>	<u>% of Untreated (Check)</u>
20	M/C-SD+H	70.6 a	452
	M/C-H	58.5 ab	375
	M/C	46.3 bc	297
	M/C+H	42.3 bc	271
	M/C-SD	40.0 cd	256
25	Check	15.6 e	100

¹ Marketable yield included all bean pods 10 cm x 0.5 cm or larger. Means followed by the same letter are not significantly different at P=0.05 according to Fisher's LSD.

As shown in Table 28, the application of *Erwinia amylovora* harpin by various methods of application resulted in an increase in the yield of marketable size snap bean pods. Treatment with methyl cellulose alone also results in an increase in bean yield but was substantially increased when combined with harpin as seed (spray dried) and foliar treatments.

Example 12 - Yield Increase in Cucumbers from Foliar Application of HP-1000™ to Cucumbers.

Cucumber seedlings and transplants were treated with foliar sprays of HP-1000™ (EDEN Bioscience, Bothell,

Washington) (*Erwinia amylovora* hypersensitive response elicitor formulation) at rates of 15, 30, or 60 $\mu\text{g/ml}$ active ingredient (a.i.). The first spray was applied when the first true leaves were fully expanded. The second application was made 10 days after the first spray. All sprays were applied using a back-pack sprayer, and an untreated control(UTC) was also included in the trial. Three days after the second application of HP-1000™, ten plants from each treatment were transplanted into randomized field plots replicated three times. This yielded a total of thirty plants per treatment. Seven days after transplanting, a third foliar spray of HP-1000™ was applied. Although severe drought followed resulting in significant water stress, a total of six harvests were made following a standard commercial harvesting pattern. The total weight of fruit harvested from each treatment is presented in Table 29. Results indicate that plants treated with HP-1000™ at rates of 15 and 30 $\mu\text{g/ml}$ yielded significantly more fruit than the UTC. Plants treated with HP-1000™ yielded a moderate yield increase. These results indicated that HP-1000™ treated plants were significantly more tolerant to drought stress conditions than untreated plants.

Table 29 - Increase yield of cucumbers after treatment with HP-1000™

Treatment	Rate ¹	Yield, ² lbs./10 plants	% above UTC
UTC	---	9.7 a	---
HP-1000™	15 $\mu\text{g/ml}$	25.4 b	161.4
HP-1000™	30 $\mu\text{g/ml}$	32.6 c	236.4
HP-1000™	60 $\mu\text{g/ml}$	11.2 a	15.9

¹Active ingredient (a.i.). ²Means followed by different letters are significantly different according to Duncan's MRT, P=0.05.

Example 13 - Yield Increase in Cotton from Treatment with HP-1000™

Cotton was planted in four, 12 x 20 foot replicate field plots in a randomized complete block (RCB) field trial. Plants were treated with HP-1000™ (EDEN Bioscience) (*Erwinia amylovora* hypersensitive response elicitor formulation), HP-1000™+Pix° (Pix° (BASF Corp., Mount Olive, N.J.) is a growth regulator applied to keep cotton plants compact in height) or Early Harvest° (Griffen Corp., Valdosta, Ga.) (a competitive growth enhancing agent). An untreated control (UTC) was also included in the trial. Using a back-pack sprayer, foliar applications were made of all treatments at three crop growth stages; first true leaves, pre-bloom, and early bloom. All fertilizers and weed control products were applied according to conventional farming practices for all treatments. The number of cotton bolls per plant ten weeks before harvest was significantly higher for the HP-1000™ treated plants compared to other treatments. By harvest, HP-1000™ treatment was shown to have a significantly increased lint yield (43%) compared to UTC (Table 30). When HP-1000™ was combined with Pix°, lint yield was increased 20% over UTC. Since Pix° is commonly applied to large acreages of cotton, this result indicates that HP-1000™ may be successfully tank-mixed with Pix°. Application of the competitive growth enhancing agent, Early Harvest° only produced a 9% increase in lint yield vs. UTC.

Table 30 - Increased lint yield from cotton after treatment with HP-1000™, HP-1000™+Pix®, or Early Harvest®.

5	-----
	Treatment Rate ¹ Lint Yield (lbs./ac) % above
	UTC
	UTC --- 942.1 ---
10	Early Harvest® 2 oz./ac. 1,077.4* 14.3
	HP-1000™+Pix® 40 µg/ml+8 oz./ac. 1,133.1* 20.4
	HP-1000™ 40 µg/ml 1,350.0* 43.3
	(*significant at P= 0.05) lsd = 122.4
15	-----

¹Rates for HP-1000™ are for active ingredient (a.i.); rates for Early Harvest® and Pix® are formulated product.

Example 14 - Yield Increase of Chinese Egg Plant from Treatment with HP-1000™

Nursery grown Chinese egg plant seedlings were sprayed once with HP-1000™ at (EDEN Bioscience) (*Erwinia amylovora* hypersensitive response elicitor formulation) 15, 30, or 60 µg/ml (a.i.), then transplanted into field plots replicated three times for each treatment. Two weeks after transplanting, a second application of HP-1000™ was made. A third and final application of HP-1000™ was applied approximately two weeks after the second spray. All sprays were applied using a back-pack sprayer; an untreated control (UTC) was also included in the trial. As the season progressed, a total of eight harvests from each treatment were made. Data from these harvests indicate that treatment with HP-1000™ resulted in greater yield of fruit per plant.

Table 31 - Increased yield for Chinese egg plant after treatment with HP-1000™.

5	-----			
	Treatment	Rate (a.i.)	Yield(lbs./plant)	% above UTC
	UTC	--	1.45	---
	HP-1000™	15 µg/ml	2.03	40.0
	HP-1000™	30 µg/ml	1.90	31.0
10	HP-1000™	60 µg/ml	1.95	34.5

15 **Example 15 - Yield Increase of Rice From Treatment with HP-1000™**

20 Rice seedlings were transplanted into field plots replicated three times, then treated with foliar sprays of HP-1000™ (EDEN Bioscience) (*Erwinia amylovora* hypersensitive response elicitor formulation) at three different rates using a back-pack sprayer. An untreated control (UTC) was also included in the trial. The first application of HP-1000™ was made one week after

25 transplanting, the second three weeks after the first. A third and final spray was made just before rice grains began to fill the heads. Results at harvest demonstrated that foliar applications of HP-1000™ at both 30 and 60 µg/ml significantly increased yield by 47 and 56%,

30 respectively (Table 32).

Table 32 - Increase yield of rice after foliar treatment with HP-1000™.

5	Treatment	Rate (a.i.)	Yield ¹ (lbs./ac.)	% above UTC
	UTC	---	3,853 a	---
	HP-1000™	15 µg/ml	5,265 ab	35.9
	HP-1000™	30 µg/ml	5,710 b	47.3
10	HP-1000™	60 µg/ml	6,043 b	56.1

15 ¹Means followed by different letters are significantly different according to Duncan's MRT, P=0.05.

Example 16 - Yield Increase of Soybeans From Treatment with HP-1000™

20 Soybeans were planted into randomized field plots replicated three times for each treatment. A back-pack sprayer was used to apply foliar sprays of HP-1000™ (EDEN Bioscience) (*Erwinia amylovora*

25 hypersensitive response elicitor formulation) and an untreated control (UTC) was also included in the trial. Three rates of HP-1000™ were applied beginning at four true leaves when plants were approximately eight inches tall. A second spray of HP-1000™ was applied ten days

30 after the first spray and a third spray ten days after the second. Plant height measured ten days after the first spray treatment indicated that application of HP-1000™ resulted in significant growth enhancement (Table 33). In addition, plants treated with HP-1000™ at

35 the rate of 60 µg/ml began to flower five days earlier than the other treatments. Approximately ten days after application of the third spray, the number of soybean pods per plant was counted from ten randomly selected plants per replication. These results indicated that the

40 growth enhancement from treatment with HP-1000™ resulted in significantly greater yield (Table 34).

Table 33 - Increased plant height of soybeans after foliar treatment with HP-1000™.

5	Treatment	Rate (a.i.)	Plant Ht. ¹ (in.)	% above UTC
	UTC	---	12.2 a	---
	HP-1000™	15 µg/ml	13.2 b	8.3
	HP-1000™	30 µg/ml	14.1 c	16.2
10	HP-1000™	60 µg/ml	14.3 c	17.3

¹Means followed by different letters are significantly different according to Duncan's MRT, P=0.05.

Table 34 - Increased pod set of soybeans after foliar treatment with HP-1000™.

20	Treatment	Rate (a.i.)	No. Pods/plant ¹	% above UTC
	UTC	---	41.1 a	---
	HP-1000™	15 µg/ml	45.4 ab	10.4
25	HP-1000™	30 µg/ml	47.4 b	15.4
	HP-1000™	60 µg/ml	48.4 b	17.7

¹Means followed by different letters are significantly different according to Duncan's MRT, P=0.05.

Example 17 - Yield Increase of Strawberries From Treatment with HP-1000™

35 Two field trials with HP-1000™ (EDEN Bioscience) (*Erwinia amylovora* hypersensitive response elicitor formulation) were conducted on two strawberry varieties, Camarosa and Selva. For each variety, a
40 randomized complete block (RCB) design was established having four replicate plots (5.33 x 10 feet) per treatment in a commercially producing strawberry field. Within each plot, strawberry plants were planted in a double row layout. An untreated control (UTC) was also
45 included in the trial. Before applications began, all plants were picked clean of any flowers and berries.

Sprays of HP-1000™ at the rate of 40 µg/ml were applied as six weekly using a back-pack sprayer. Just prior to application of each spray, all ripe fruit from each treatment was harvested, weighed, and graded according to commercial standards. Within three weeks of the first application of HP-1000™ to *Selva* strawberry plants, growth enhancement was discernible as visibly greater above-ground biomass and a more vigorous, greener and healthier appearance. After six harvests (i.e. the scheduled life-span for these plants), all yield data were summed and analyzed. For the *Camarosa* variety, yield of marketable fruit from HP-1000™ treated plants was significantly increased (27%) over the UTC when averaged over the last four pickings (Table 35). Significant differences between treatments were not apparent for this variety for the first two pickings. The *Selva* variety was more responsive to the growth enhancing effects from treatment with HP-1000™; *Selva* strawberry plants yielded a statistically significant 64% more marketable fruit vs. the UTC when averaged over six pickings (Table 35).

Table 35 - Increased yield of strawberries after foliar treatment with HP-1000™.

Treatment	Rate (a.i.)	Yield ¹ (lbs./rep)	% above
UTC			
Variety: <i>Camarosa</i>			
UTC	---	1.71 a	---
HP-1000™	40 µg/ml	2.17 b	27
Variety: <i>Selva</i>			
UTC	---	0.88 a	---
HP-1000™	40 µg/ml	1.44 b	64

¹Means followed by different letters are significantly different according to Duncan's MRT, P=0.05.

Example 18 - Earlier Maturity and Increased Yield of Tomatoes from Treatment with HP-1000™

Fresh market tomatoes (var. *Solar Set*) were
5 grown in plots (2 x 30 feet) replicated 5 times in a
randomized complete block (RCB) field trial within a
commercial tomato production field. Treatments included
HP-1000™ (EDEN Bioscience) (*Erwinia amylovora*
hypersensitive response elicitor formulation), an
10 experimental competitive product (Actigard™ (Novartis,
Greensboro, N.C.)) and a chemical standard (Kocide®
(Griffen Corp., Valdosta, GA)) + Maneb® (DuPont
Agricultural Products, Wilmington, D.E.)) for disease
control. The initial application of HP-1000™ was made as
15 a 50 ml drench (of 30 µg/ml a.i.) poured directly over
the seedling immediately after transplanting.
Thereafter, eleven weekly foliar sprays were applied
using a back-pack sprayer. The first harvest from all
treatments was made approximately six weeks after
20 transplanting and only fully red, ripe tomatoes were
harvested from each treatment. Results indicated that
HP-1000™ treated plants had a significantly greater
amount of tomatoes ready for the first harvest
(Table 36). The tomatoes harvested from the HP-1000™
25 treated plants were estimated to be 10-14 days ahead
other treatments.

Table 36 - Increased yield of tomatoes at first harvest after foliar treatment with of HP-1000™.

5	Treatment	Rate (a.i.) ¹	Yield ² (lbs./rep)	% above
	UTC			
	UTC	---	0.61 a	---
	HP-1000™	30 µg/ml	2.87 b	375
10	Actigard™	14 g/ac	0.45 a	-25.1
	Kocide®	2 lbs./ac.	0.31 a	-49.1
	Maneb®	1 lb./ac		

¹Rates for Kocide® and Maneb® are for formulated product. ²Means followed by different letters are significantly different according to Duncan's MRT, P=0.05.

Example 19 - Earlier Flowering and Growth Enhancement of Strawberries From Treatment with HP-1000™ When Planted in Non-fumigated Soil.

Strawberry plants ("plugs" and "bare-root"), cv. *Commander* were transplanted into plots (2 x 30 feet) replicated 5 times in a randomized complete block field trial. Approximately sixty individual plants were transplanted into each replicate. Treatments applied in this field trial are listed below:

	<u>Treatment</u>	<u>Application method</u>
35	HP-1000™ (plug plants)	50-ml drench solution of HP-1000™ (EDEN Bioscience) (<i>Erwinia amylovora</i> hypersensitive response elicitor formulation) at 40 µg/ml (a.i.) poured directly over the individual plants immediately after transplanting into non-fumigated soil ¹ , followed by foliar applications of HP-1000™ at 40 µg/ml every 14 days.
40		
45	HP-1000™ 40 (bare-root plants)	root soak in solution of HP-1000™ at µg/ml (a.i.) for 1 hour, immediately before transplanting into non-fumigated

soil,¹ followed by foliar applications of HP-1000™ at 40 µg/ml every 14 days.

- 5 methyl bromide/
chlorpicrin
75/25 soil fumigation at 300 lbs./ac via
injection prior to transplanting, no
HP-1000™ treatments applied.
- 10 Telone/chlorpicrin
70/30 soil fumigation at 45 gal./ac via
injection prior to transplanting, no
HP-1000™ treatments applied.
- untreated control no fumigation, no HP-1000™ treatments
(UTC)
- 15 ¹Non-fumigated soil had been cropped to vetch for the two previous
years.
- 20 Transplanting was done in late fall when cool weather
tended to slow plant growth. Two weeks after
transplanting, the first foliar application of HP-1000™
was made at 40 µg/ml (a.i.) with a back-pack sprayer.
Three weeks after transplanting, preliminary results were
- 25 gathered comparing HP-1000™ treatment against methyl
bromide and UTC by counting the number of flowers on all
strawberry "plug" plants in each replication. Since
flowering had not yet occurred in the "bare-root" plants,
each plant in replicates for this treatment was assessed
- 30 for early leaf growth by measuring the distance from leaf
tip to stem on the middle leaf of 3-leaf cluster.
Results (Tables 37 and 38) indicated that treatment with
HP-1000™ provided early enhanced flower growth and leaf
size for "plug" and "bare-root" strawberry plants,
- 35 respectively.

Table 37 - Earlier flowering of "plug" strawberry transplants after foliar treatment with HP-1000™.

5	-----				
	Treatment	Rate (a.i.)	No. flowers/rep ¹	% above	
	UTC				
	UTC	---	2.0a	---	
10	HP-1000™	40 µg/ml	7.5 b	275	
	Methyl bromide/ chlorpicrin	300 lbs./ac	5.3 b	163	

¹Means followed by different letters are significantly different according to Duncan's MRT, P=0.05.

Table 38 - Increased leaf growth in "bare-root" strawberry transplants after foliar treatment with HP-1000™.

25	-----				
	Treatment	Rate (a.i.)	Leaf length ¹ (in.)	% above	
	UTC				
	UTC	---	1.26 a	---	
30	HP-1000™	40 µg/ml	1.81 b	44	

¹Means followed by different letters are significantly different according to Duncan's MRT, P=0.05.

Example 20 - Early Growth Enhancement of Jalapeño Peppers from Application of HP-1000™

40 Jalapeño pepper (cv. Mittlya) transplants were treated with a root drench of HP-1000 (EDEN Bioscience) (*Erwinia amylovora* hypersensitive response elicitor formulation) (30 µg/ml a.i.) for 1 hour, then

45 transplanted into randomized field plots replicated four times. An untreated control (UTC) was also included. Beginning 14 days after transplanting, treated plants received three foliar sprays of HP-1000™ at 14 day

intervals using a back-pack sprayer. One week after the third application of HP-1000™ (54 days after transplanting), plant height was measured from four randomly selected plants per replication. Results from these measurements indicated that the HP-1000™ treated plants were approximately 26% taller than the UTC plants (Table 39). In addition, the number of buds, flowers or fruit on each plant was counted. These results indicated that the HP-1000™ treated plants had over 61% more flowers, fruit or buds compared to UTC plants (Table 40).

Table 39 - Increased plant height in Jalapeño peppers after treatment with HP-1000™.

15	-----			
	Treatment	Rate (a.i.)	Plant Ht.(in.) ¹	% above UTC
	UTC	---	a7.0	---
20	HP-1000™	30 µg/ml	8.6 b	23.6

¹Means followed by different letters are significantly different according to Duncan's MRT, P=0.05.

Table 40 - Increased number of flowers, fruit or buds in Jalapeño peppers after treatment with HP-1000™.

Treatment	Rate (a.i.)	No. flowers, fruit or buds/plant ¹	% above
UTC	---	20.6 a	---
HP-1000™	30 µg/ml	12.8 b	61.3

¹Means followed by different letters are significantly different according to Duncan's MRT, P=0.05.

Example 21 - Growth Enhancement of Tobacco from
Application of HP-1000™

Tobacco seedlings were transplanted into
5 randomized field plots replicated three times. A foliar
spray of HP-1000™ (EDEN Bioscience) (*Erwinia amylovora*
hypersensitive response elicitor formulation) was applied
after transplanting at one of three rates: 15, 30, or 60
μg/ml a.i. Sixty days later, a second foliar application
10 of HP-1000 was made. Two days after the second
application, plant height, number of leaves per plant,
and the leaf size (area) were measured from ten, randomly
selected plants per treatment. Results from these
measurements indicated treatment with HP-1000™ enhanced
15 tobacco plant growth significantly (Tables 41, 42, and
43). Plant height was increased by 6-13%, while plants
treated with HP-1000™ at 30 and 60 μg/ml averaged just
over 1 more leaf per plant than UTC. Most significantly,
however, treatment with HP-1000™ at 15, 30, and 60 μg/ml
20 resulted in corresponding increases in leaf area.
Tobacco plants with an extra leaf per plant and an
increase in average leaf size (area) represent a
commercially significant response.

25 Table 41 - Increased plant height in tobacco after
treatment with HP-1000™.

30

Treatment	Rate (a.i.)	Plant Ht. (cm)	% above UTC
UTC	---	72.0	---
HP-1000™	15 μg/ml	76.4	5.3
HP-1000™	30 μg/ml	79.2	9.0
35 HP-1000™	60 μg/ml	81.3	6.9

40

Table 42 - Increased number of tobacco leaves per plant after treatment with HP-1000™.

5	Treatment	Rate (a.i.)	Leaves/plant ¹	% above UTC
	UTC	---	16.8	---
	HP-1000™	15 µg/ml	17.4	3.6
	HP-1000™	30 µg/ml	18.1	7.7
10	HP-1000™	60 µg/ml	17.9	6.5

Table 43 - Increased leaf area in tobacco after treatment with HP-1000™.

15	20	Treatment	Rate (a.i.)	Leaf area (cm ²)	% above UTC
		UTC	---	1,246	---
		HP-1000™	15 µg/ml	1,441	16
		HP-1000™	30 µg/ml	1,543	24
	25	HP-1000™	60 µg/ml	1,649	32

Example 22 - Growth Enhancement of Winter Wheat from Application of HP-1000™

Winter wheat seed was "dusted" with dry HP-1000™ (EDEN Bioscience) (*Erwinia amylovora* hypersensitive response elicitor formulation) powder at the rate of 3 ounces of formulated product (3% a.i.) per 100 lbs. seed, then planted using conventional seeding equipment into randomized test plots 11.7 feet by 100 feet long. Additional treatments included a seed "dusting" with HP-1000™ powder (3% a.i.) at 1 oz. formulated product per 100 lbs. seed, a seed-soak in a solution of HP-1000™ at a concentration of 20 µg/ml, a.i., for four hours, then air-dried before planting, a standard chemical (Dividend®) fungicide "dusting", and an untreated control (UTC). Eight days after planting,

HP-1000™ treated seeds began to emerge, whereas the UTC and chemical standard-treated seed did not emerge until approximately 14 days after planting, the normal time expected. At 41 days after planting, seedlings were
5 removed from the ground and evaluated. Root mass for wheat treated with HP-1000™ as a "dusting" at 3 oz./100 lb. was visually inspected and judged to be approximately twice as great as any of the other treatments.

Following the field trial, a greenhouse
10 experiment was designed to gain confirmation of these results. Treatments included wheat seed dusted with dry HP-1000™ (10% a.i.) at a rate of 3 ounces per 100 lbs. of seed, seed soaking of HP-1000™ in solution concentration of 20 mg/ml for four hours before planting, and an
15 untreated control (UTC). Wheat seeds from each treatment were planted at the rate of 25 seeds per pot, with five pots serving as replicates for each treatment. Fifteen days after planting, ten randomly selected seedlings from each treatment pot were removed, carefully cleaned, and
20 measured for root length. Since the above-ground portion of individual seedlings did not exhibit any treatment effect, increased root growth from treatment with HP-1000™ did not influence the selection of samples. The increase in root growth from either HP-1000™ treatment
25 was significantly greater than UTC (Table 49); however, the seed dusting treatment appeared to give slightly better results.

Table 44 - Increased root growth in wheat seedlings after treatment with HP-1000™.

5	Treatment	Rate	Root length. (cm) ¹	% above UTC
	UTC	---	35.6 a	---
	HP-1000™			
	(dusting)	3 oz./100 lbs.	41.0 b	17.4
10	HP-1000™			
	(soaking)	20 µg/ml	40.8 b	14.6

15 ¹Means followed by different letters are significantly different according to Duncan's MRT, P=0.05.

20 **Example 23 - Growth Enhancement of Cucumbers from Application of HP-1000™**

A field trial of commercially produced cucumbers consisted of four treatments, HP-1000™ (EDEN Bioscience) (*Erwinia amylovora* hypersensitive response elicitor formulation) at two rates (20 or 40 µg/ml), a chemical standard for disease control (Bravo® (Zeneca Ag Products, Wilmington, Del.) +Maneb®) and an untreated control (UTC). Each treatment was replicated four times in 3 x 75 foot plots with a plant spacing of approximately 2 feet for each treatment. Foliar sprays of HP-1000™ were applied beginning at first true leaf and repeated at 14 day intervals until the last harvest for a total of six applications. The standard fungicide mix was applied every seven days or sooner if conditions warranted. Commercial harvesting began approximately two months after first application of HP-1000™ (after five sprays of HP-1000™ had been applied), and a final harvest was made approximately 14 days after the first harvest.

Results from the first harvest indicated that treatment with HP-1000™ enhanced the average cucumber yield by increasing the total number of cucumbers

harvested and not the average weight of individual cucumbers (Tables 45-47). The same trend was noted at the final harvest (Tables 48-49). It was commercially important that the yield increase resulting from treatment with HP-1000™ was not achieved by significantly increasing average cucumber size.

Table 45 - Increased cucumber yield after treatment with HP-1000™, first harvest.

Treatment	Rate (a.i.)	Yield/trt ¹ (kg.)	% above UTC
UTC	---	10.0 a	---
Bravo+Maneb	label	10.8 a	8.4
HP-1000™	20 µg/ml	12.3 ab	22.8
HP-1000™	40 µg/ml	13.8 b	38.0

¹Means followed by different letters are significantly different according to Duncan's MRT, P=0.05.

Table 46 - Increased number of fruit in cucumbers after treatment with HP-1000™, first harvest.

Treatment	Rate (a.i.)	No. fruit/trt ¹	% above UTC
UTC	---	24.5 a	---
Bravo+Maneb	label	27.6 ab	12.8
HP-1000™	20 µg/ml	31.2 b	27.0
HP-1000™	40 µg/ml	34.3 b	39.8

¹Means followed by different letters are significantly different according to Duncan's MRT, P=0.05.

Table 47 - Average weight of cucumbers after treatment with HP-1000™, first harvest.

	Treatment	Rate (a.i.)	Weight/fruit(g)	% change vs.
	UTC	---	406	---
	Bravo+Maneb	label	390	-4
10	HP-1000™	20 µg/ml	395	-3
	HP-1000™	40 µg/ml	403	-1

15

Table 48 - Increased cucumber yield after treatment with HP-1000™, third harvest.

	Treatment	Rate (a.i.)	Yield/trt ¹ (kg.)	% above UTC
	UTC	---	17.5 a	---
	Bravo+Maneb	label	14.0 b	-20.1
25	HP-1000™	20 µg/ml	20.1 a	15.3
	HP-1000™	40 µg/ml	20.2 a	15.6

30 'Means followed by different letters are significantly different according to Duncan's MRT, P=0.05.

35 Table 49 - Increased number of fruit in cucumbers after treatment with HP-1000™, third harvest.

	Treatment	Rate (a.i.)	No. fruit/trt ¹	% change vs.
	UTC	---	68.8 ab	---
	Bravo+Maneb	label	60.0 a	-12.7
	HP-1000™	20 µg/ml	82.3 b	19.6
45	HP-1000™	40 µg/ml	85.3 b	24.0

50 'Means followed by different letters are significantly different according to Duncan's MRT, P=0.05.

Table 50 - Average weight of cucumbers after treatment with HP-1000™, third harvest.

5	Treatment	Rate (a.i.)	Weight/fruit(g)	% change vs.
	UTC			
	UTC	---	255	---
	Bravo+Maneb	label	232	-9
10	HP-1000™	20 µg/ml	247	-3
	HP-1000™	40 µg/ml	237	-7

15 Example 24 - Harpin_{pss} from *Pseudomonas syringae* pv *syringae* Induces Growth Enhancement in Tomato

20 To test if harpin_{pss} (i.e. the hypersensitive response elicitor from *Pseudomonas syringae* pv *syringae*) (He, S. Y., et al., "*Pseudomonas syringae* pv *syringae* Harpin_{pss}. A Protein that is Secreted via the Hrp Pathway and Elicits the Hypersensitive Response in Plants," Cell 25 73:1255-66 (1993), which is hereby incorporated by reference) also stimulates plant growth, tomato seeds (Marglobe variety) were sowed in 8 inches pots with artificial soil. 10 days after sowing, the seedlings were transplanted into individual pots. Throughout the 30 experiment, fertilizer, irrigation of water, temperature, and soil moisture were maintained uniformly among plants. 16 days after transplanting, the initial plant height was measured and the first application of harpin_{pss} was made, this is referred to as day 0. A second application was 35 made on day 15. Additional growth data was collected on day 10 and day 30. The final data collection on day 30 included both plant height and fresh weight.

The harpin_{pss} used for application during the experiment was produced by fermenting *E. coli* DH5 40 containing the plasmid with the gene encoding harpin_{pss} (i.e. *hrpZ*). The cells were harvested, resuspended in 5 mM potassium phosphate buffer, and disrupted by

sonication. The sonicated material was boiled for 5 minutes and then centrifugated for 10 min. at 10,000 rpm. The supernatant was considered as Cell-Free Elicitor Preparation (CFEP). 20 and 50 µg/ml harpin_{pss} solution was made with the same buffer used to make cell suspension. CFEP prepared from the same strain containing the same plasmid but without *hrpZ* gene was used as the material for control treatment.

The wetting agent, Pinene II (Drexel Chemical Co., Memphis, Tenn.) was added to the harpin_{pss} solution at the concentration of 0.1%, then harpin_{pss} was sprayed onto tomato plant until there was run off.

Table 51 shows that there was a significant difference between the harpin_{pss} treatment groups and the control group. Harpin_{pss} treated tomato increased more than 10% in height. The data supports the claim that harpin_{pss} does act similar to the hypersensitive response elicitor from *Erwinia amylovora*, in that when applied to tomato and many other species of plants, there is a growth enhancement effect. In addition to a significant increase of tomato height harpin_{pss}-treated tomato had more biomass, big leaves, early flower setting, and over all healthier appearance.

Table 51 - Harpin_{pss} enhances the growth of tomato plant

Treatment	Plant Height (cm ¹)			
	Day 0	Day 10	Day 30	
CFEP Control	8.5 ² (0.87) a ³	23.9 (1.90) a	68.2	(8.60) a
Harpinpss 20 µg/ml	8.8 (0.98) a	27.3 (1.75) b	74.2	(6.38) b
Harpinpss 50 µg/ml	8.8 (1.13) a	26.8 (2.31) b	75.4	6.30) b

¹Plant height was measured to the nearest 0.5 cm. Day 0 refers to the day the initial plant heights were recorded and the first application was made.

